

# LowSalt project

**WP1**

food matrix and  
interactions

**Supervisors**

Achim Kohler / Nils Kristian Afseth / Ragni Ofstad

**PhD student:**

Nebojsa Perisic



## overall aim of the project

- to identify innovations that can reduce the NaCl level to 50 % of the current content of commercial products such as minced fish and meat, cured ham or smoked salmon.

## NaCl reduction & substitution: focus on effects on:

- protein structural changes
- hydration properties of proteins – water interactions
- hydration properties of meat – WHC
- sensory properties / texture

## methodology

- FTIR microscopy : existing platform + further development
- Raman microscopy
- NIR spectroscopy
- multivariate analysis



## M 1.1

- new methodology to study water and protein structures in a model system

## M 1.2

- salt substitutes and/or water-binders which mimic protein-water matrix as obtained with high NaCl-concentrations ( $> 3\%$ ) in a model system are identified

## M 1.3

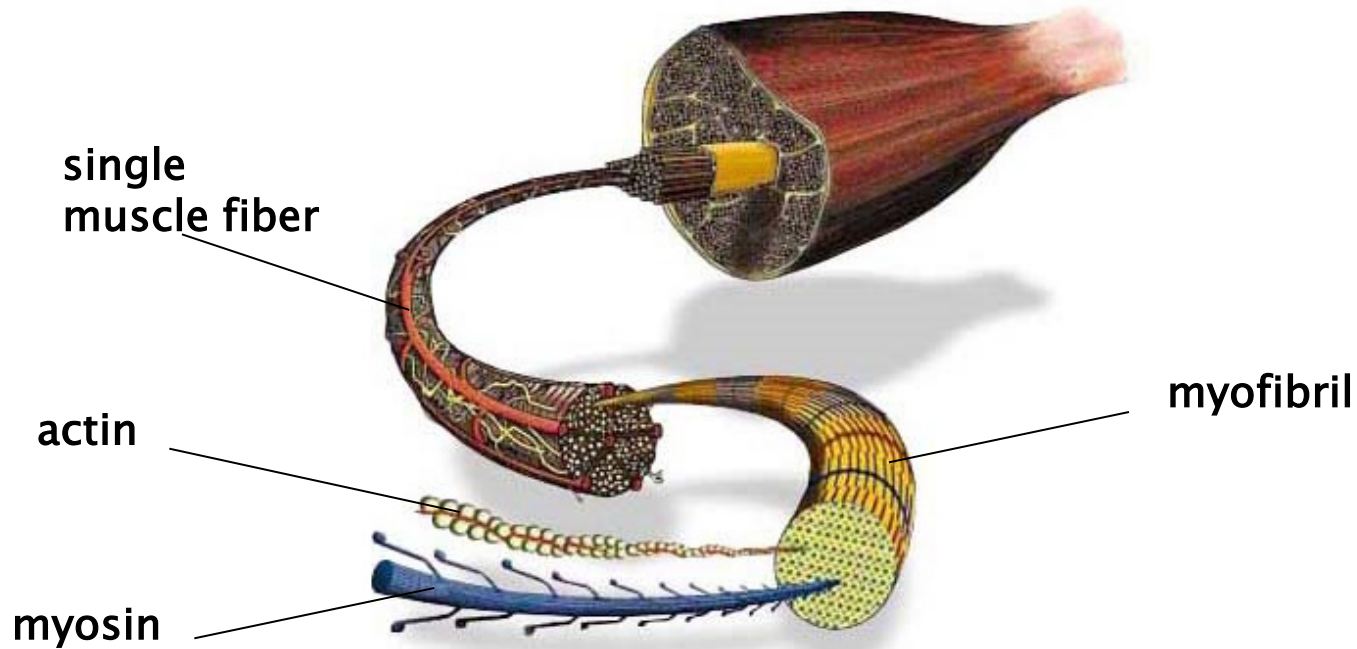
- salt substitutes and/or water-binders which mimic high salt-protein-water matrix in minced fish and meat systems are identified

## M 1.4

- Testing of selected salt substitutes and/or water-binders in minced fish and meat products completed

## M 1.5

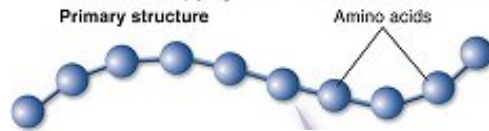
- Testing of selected salt substitutes and/or low salt technologies in cured ham completed



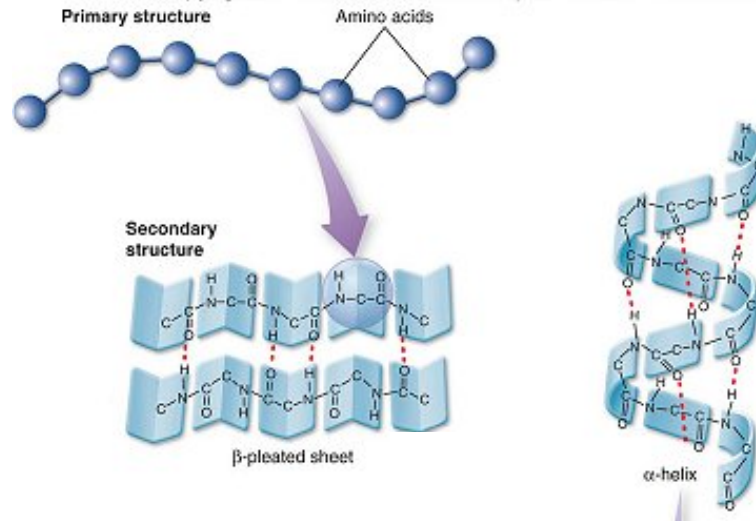
## meat

water (~75%)  
protein (~19%)  
fat (~3%)  
soluble non-protein substances (~3.5%).

- proteins
- actin
  - myosin
- } main constituents
- $\alpha$ -actinin
  - $\beta$ -actinin
  - tropomyosin
  - troponin
  - C protein
  - M line protein
- } accessory proteins



- order of the individual building blocks
- AA connected by peptide bond



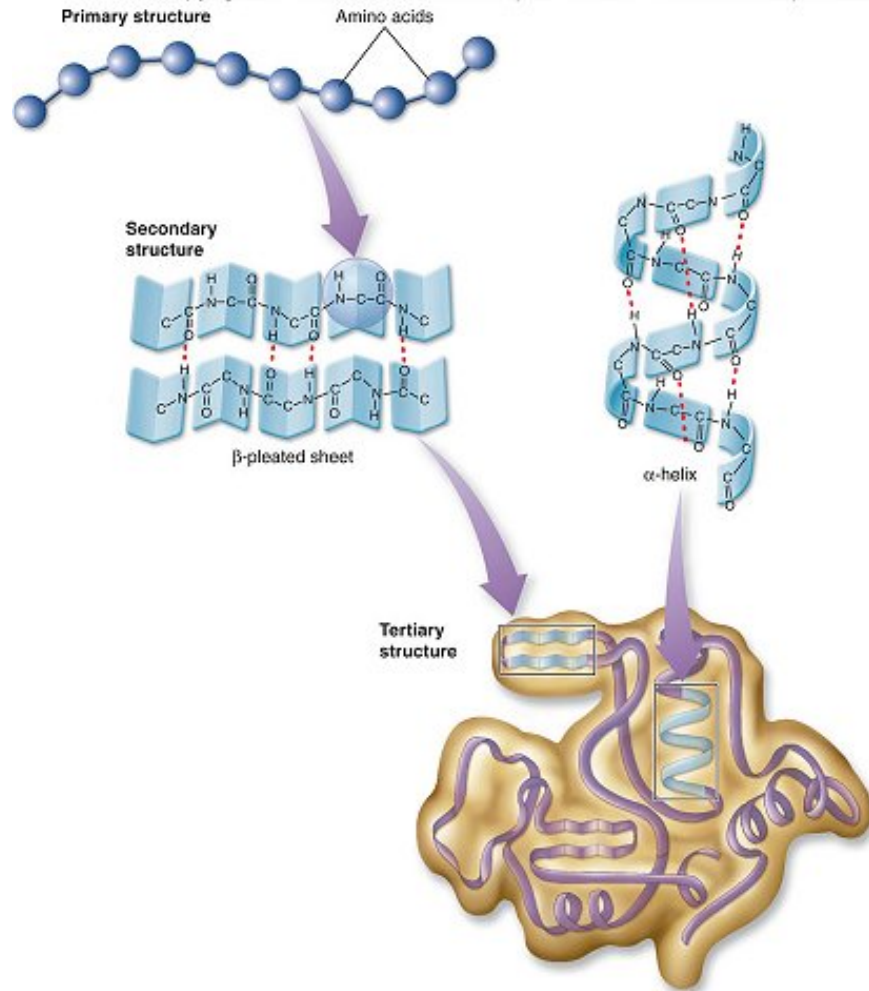
## MOLECULAR STRUCTURE

Primary (sequence)



Secondary (local folding)

- how these building blocks relate to each other
- stabilised by several interactions



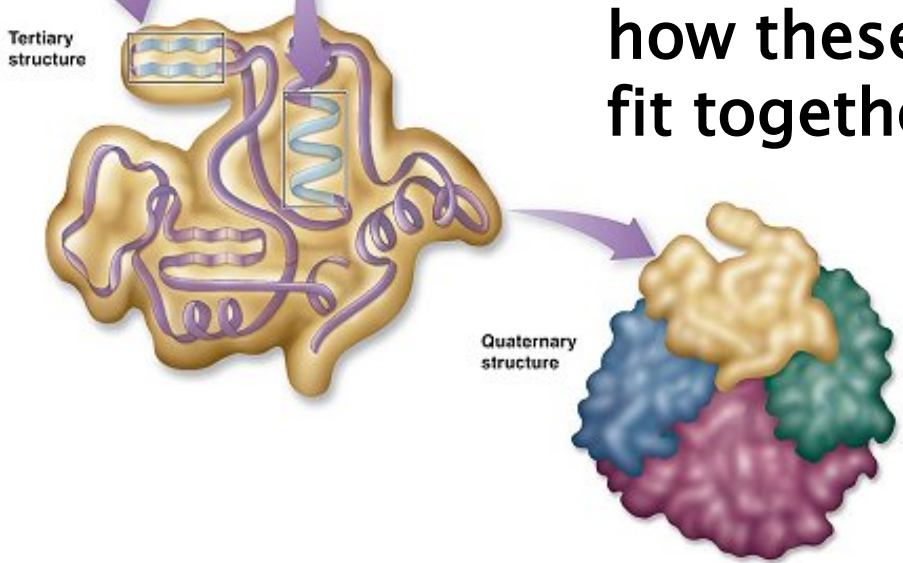
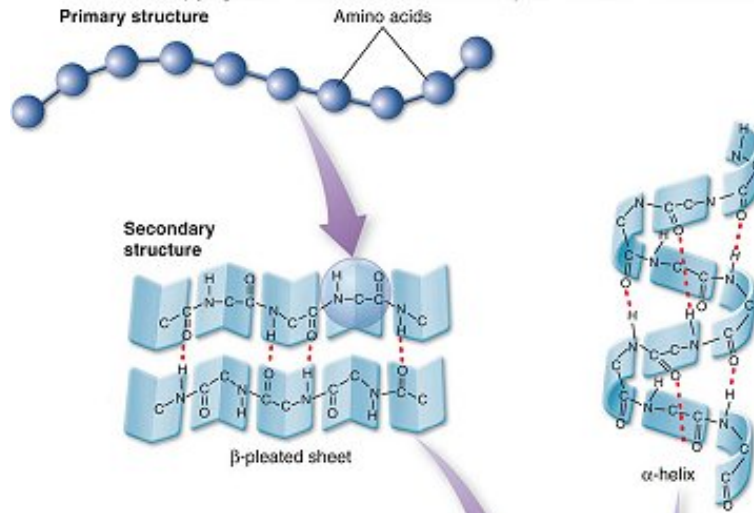
## MOLECULAR STRUCTURE

Primary (sequence)

Secondary (local folding)

Tertiary (long-range folding)

- how these fold in a 3D unit
- inter-relation between 2<sup>nd</sup> structures



## MOLECULAR STRUCTURE

Primary (sequence)

Secondary (local folding)

Tertiary (long-range folding)

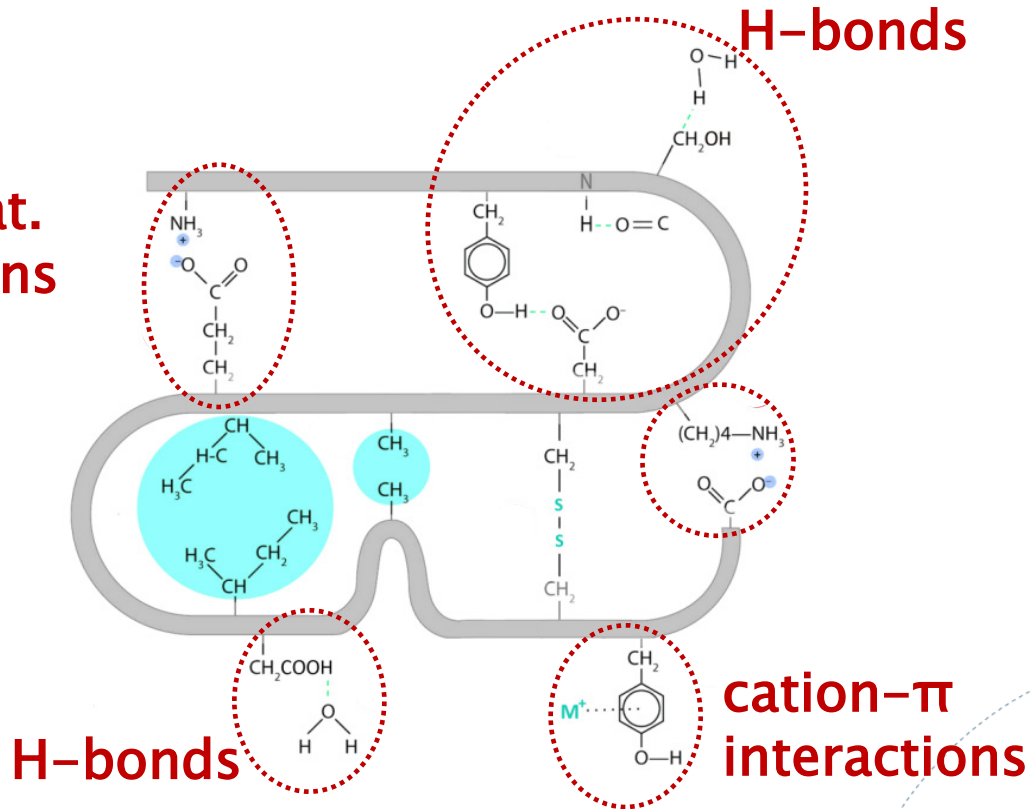
Quaternary (multimeric organization)

Supramolecular (large-scale assemblies)

## how these units fit together



electrostat.  
interactions

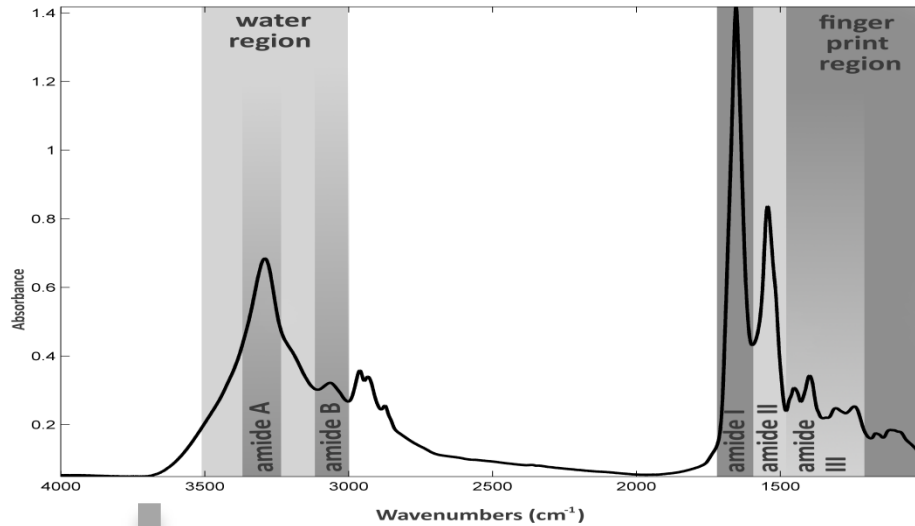


protein structure  
stabilising forces  
affected by salts

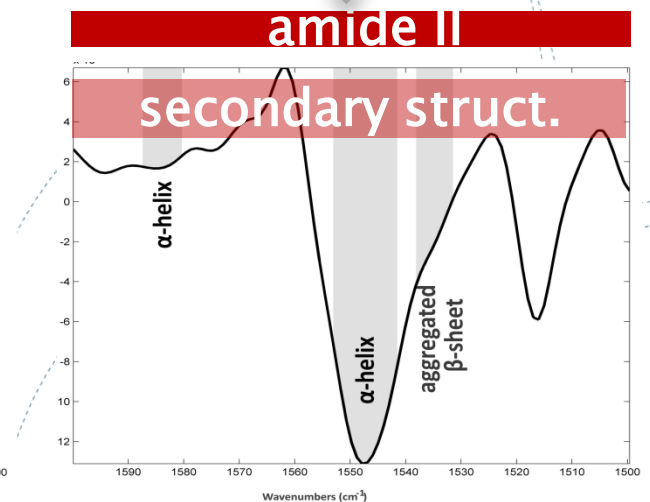
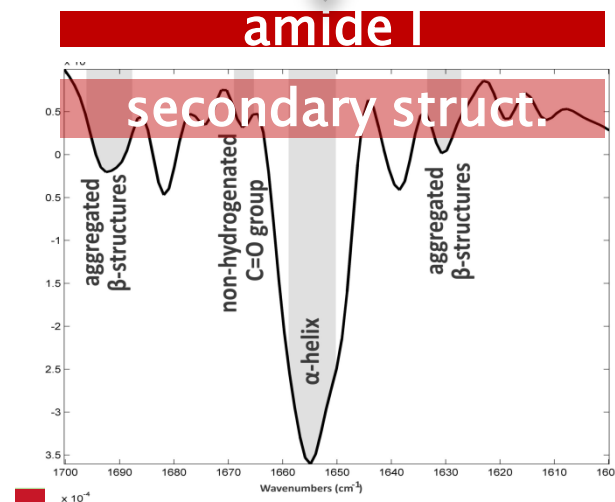
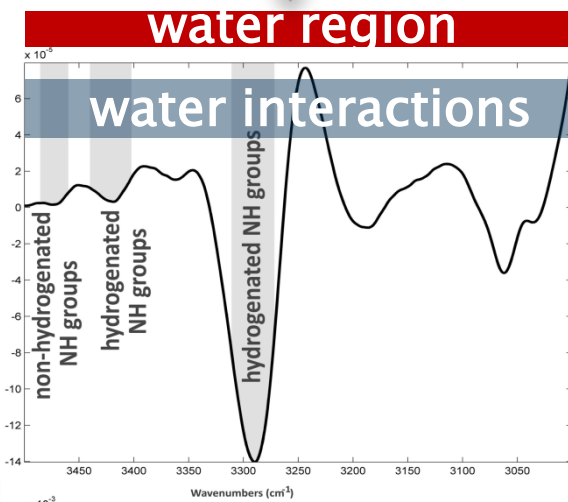
protein structure  
stabilising forces

# background

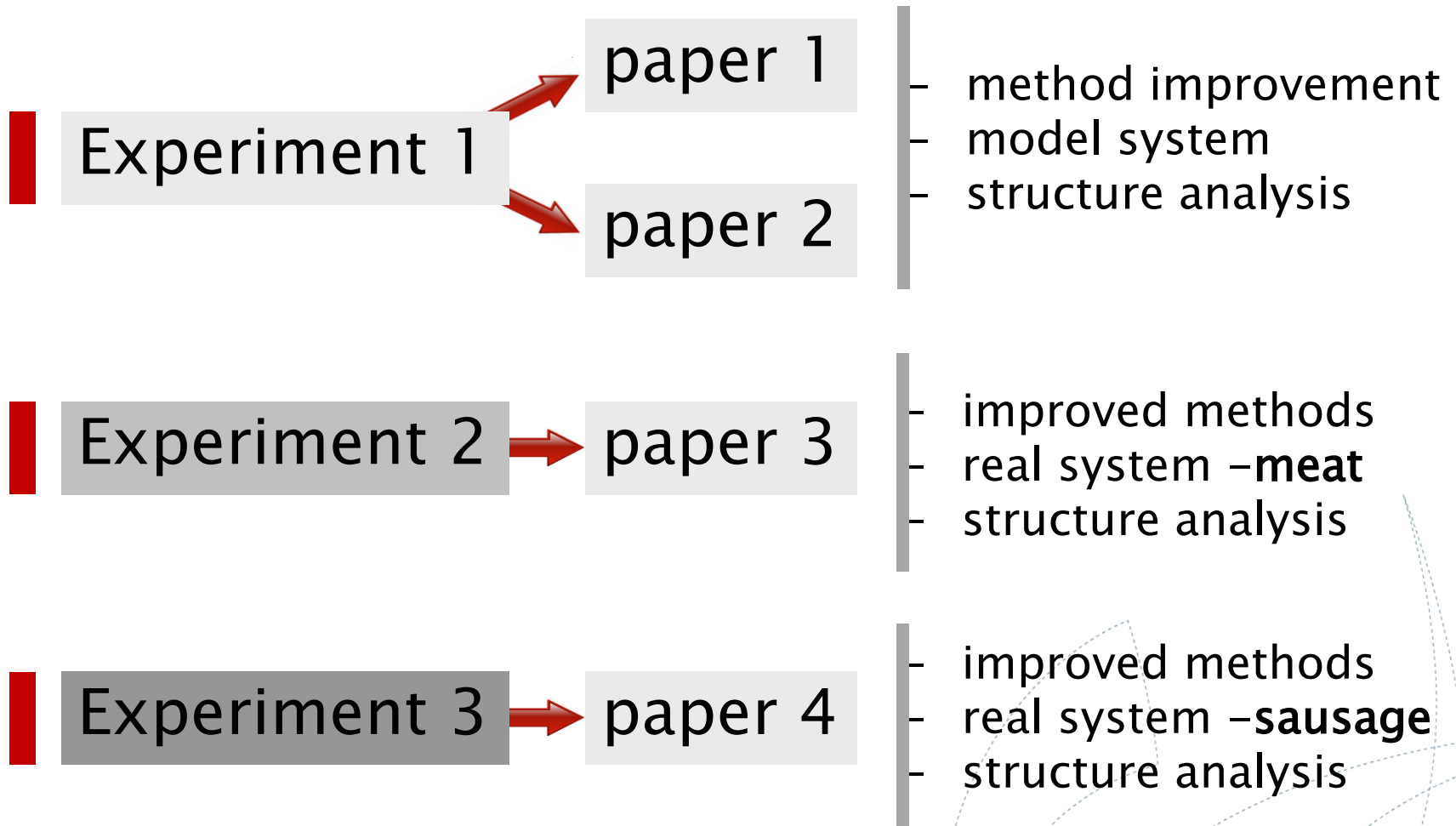
# protein structure & methodology



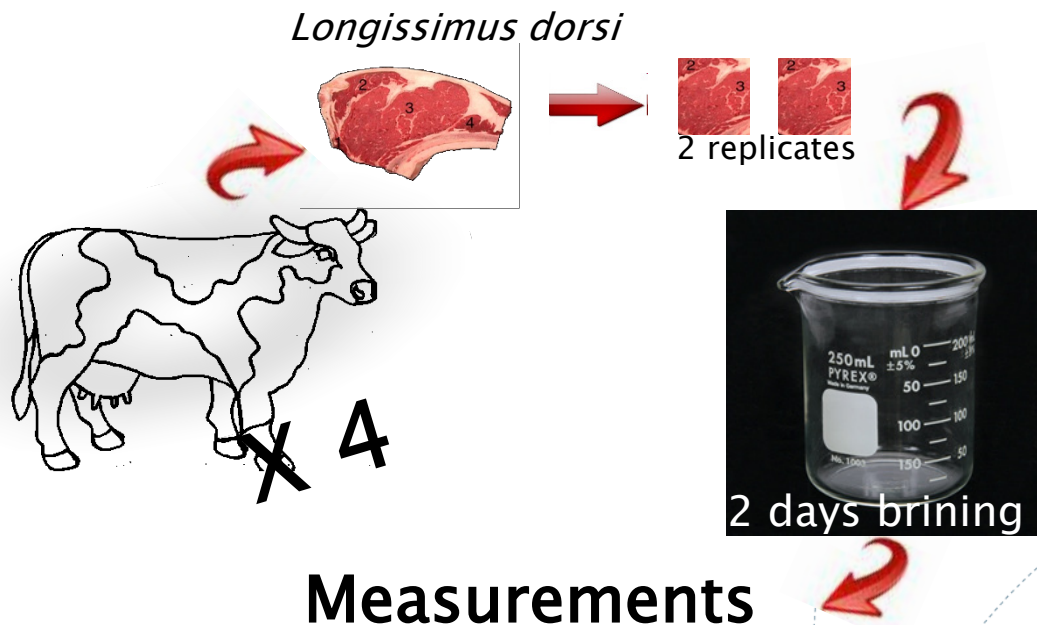
second derivative



# outline – publications



- 3 salts – NaCl / KCl / MgSO<sub>4</sub>
- 3 concentrations – 1,5 % / 6 % / 9 %
- Experiment 1
- 4 animals (beef)
- 3 spectroscopic methods – FTIR, Raman, NIR



## Monitoring Protein Structural Changes and Hydration in Bovine Meat Tissue Due to Salt Substitutes by Fourier Transform Infrared (FTIR) Microspectroscopy

### Journal of Agricultural and Food Chemistry

Perisic, N., Afseth, N. K., Ofstad, R., & Kohler, A. Monitoring Protein Structural Changes and Hydration in Bovine Meat Tissue Due to Salt Substitutes by Fourier Transform Infrared (FTIR) Microspectroscopy. *Journal of Agricultural and Food Chemistry*, 2011, 59(18), 10052–10061.

## Step 1: literature overview– band assignement

Table 1. Band Positions and Assignments for the Amide I, Amide II, and Water Regions According to the Literature and Our Previous Work

region	freq (cm <sup>-1</sup> )	tentative assignment <sup>a</sup>
amide I 1700–1600 cm <sup>-1</sup> (80% C=O stretch, 10% C–N stretch, 10% N–H bend)	1693	aggregated $\beta$ -sheet structures (sideband of 1631 cm <sup>-1</sup> band), <sup>32–35</sup> M/P
	1682	native (parallel/antiparallel) $\beta$ -sheet structures, <sup>20,32,33,37</sup> M/P/T
	1674	tentatively assigned to turns, <sup>22</sup> M/P
	1667	nonhydrogenated C=O group, internal random coil segments that are not involved in H-bonding, <sup>33,35</sup> M/P
	1660	loop structures/ $\alpha$ -helical structures, <sup>16,43,51</sup> M/P
	1655	$\alpha$ -helical structures, C=O stretching vibrations originating from $\alpha$ -helical structures in the myofibrillar proteins, <sup>20,32,33,37</sup> M/P/T or water vibrations, <sup>36,38</sup> P
	1638	water deformation mode in liquid water <sup>19,52</sup> P or native (parallel/antiparallel) $\beta$ -sheet structures, <sup>22,33</sup> M/P/T
	1631	aggregated $\beta$ -sheet structures, <sup>32–35</sup> M/P
	1618	aggregated $\beta$ -sheet structures, <sup>16,43,51</sup> M
	1611	tyrosine amino acid side chain vibrations <sup>33,40</sup> or aggregated strands, <sup>41</sup> M/P
amide II 1600–1500 cm <sup>-1</sup> (60% N–H bend, 40% C–N stretch)	1594	not assigned
	1584	$\alpha$ -helical structures, <sup>33</sup> M
	1575	amide II unspecified, <sup>33</sup> M
	1567	residue and/or possibly aggregated $\beta$ -sheet structures, <sup>33,42</sup> M
	1547	$\alpha$ -helical structures, <sup>33</sup> M/P
	1537	possibly aggregated $\beta$ -sheet structures, <sup>33</sup> M
1516	possibly tyrosine, <sup>33</sup> M/P	
water region 3500–3000 cm <sup>-1</sup> (N–H stretching, C–N–H stretching vibration, O–H stretching vibration)	3473	nonhydrogenated N–H groups, <sup>19,22,39</sup> P
	3420	hydrogenated N–H groups or O–H stretching band, <sup>21,22,39,53</sup> T/P
	3361	companion band of 1530 cm <sup>-1</sup> band, in solution occurring at 3345 cm <sup>-1</sup> , and/or N–H stretching band, <sup>21,39</sup> T/P
	3290	N–H stretching band/amide A, <sup>21,22,39,42</sup> T/P, or hydrogen-bonded NH groups <sup>19</sup> P
	3190	not assigned
	3063	N–H stretch/amide B/amide II overtone/amide II combination mode in $\beta$ -sheet structures, <sup>22,39</sup> T/P
3035	not assigned	

<sup>a</sup>M, obtained in real meat system; P, obtained in pure protein or polypeptide model system; T, obtained by theoretical calculation and/or prediction.

secondary  
structure

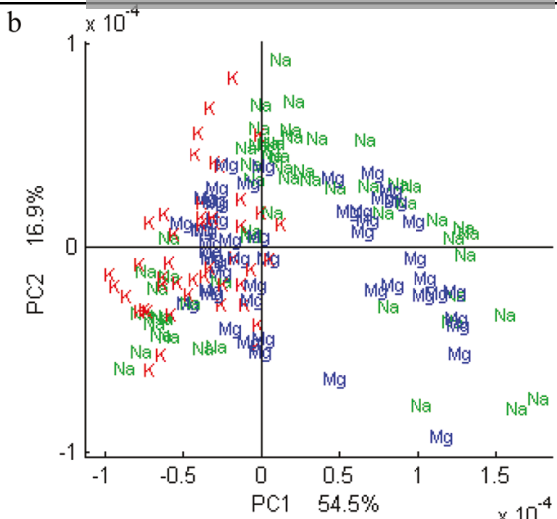
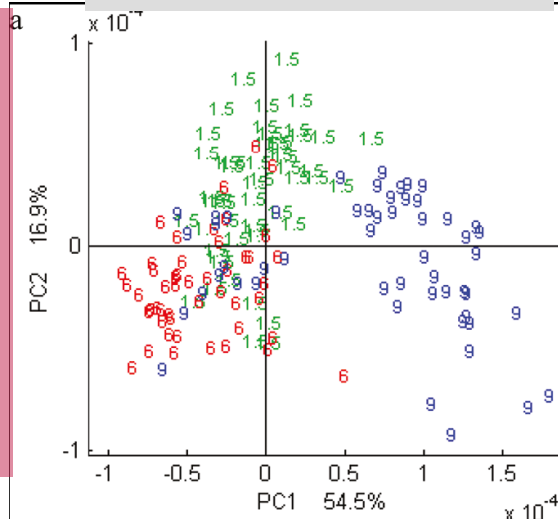
water  
interact

## Step 2: PCA of separate regions

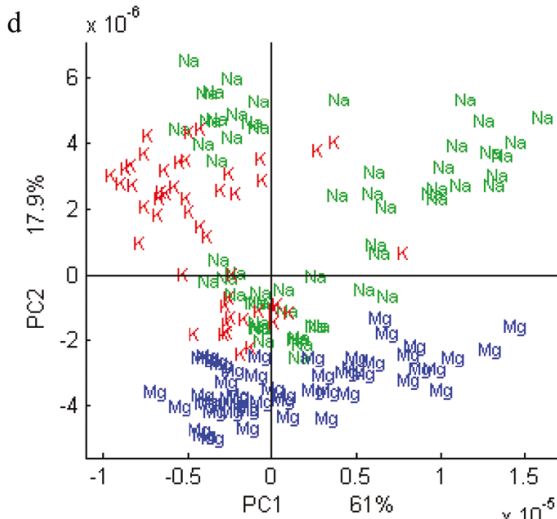
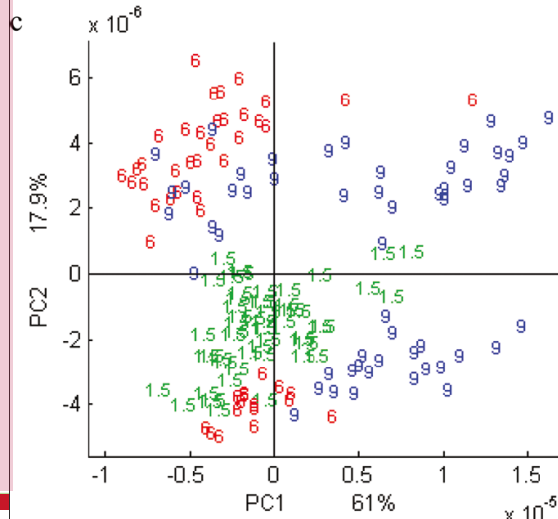
concentration

salt type

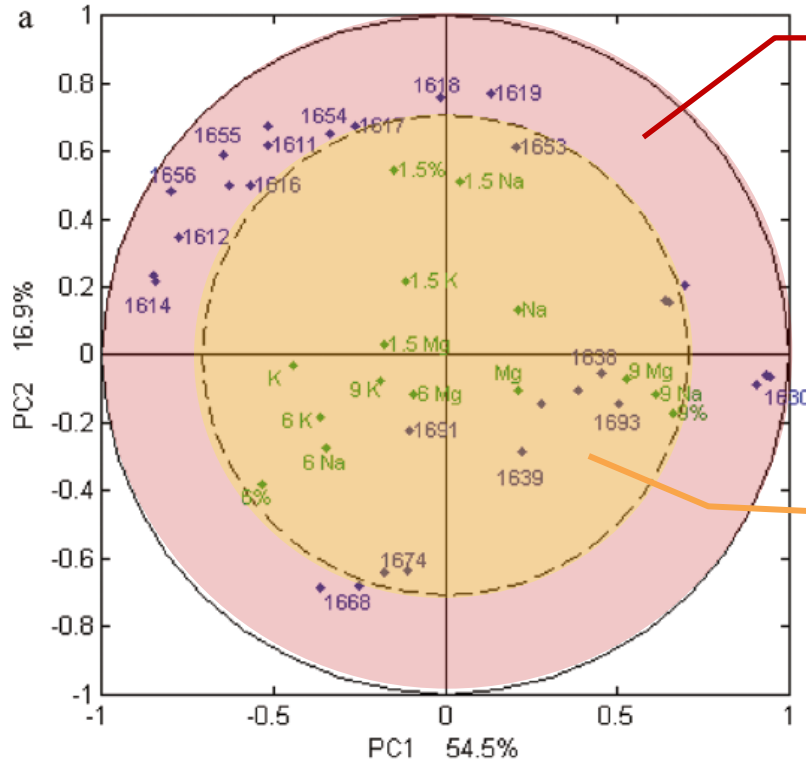
protein secondary structure



protein water interactions



## Step 2: PCA of separate regions correlation loading plots: relating to design variables



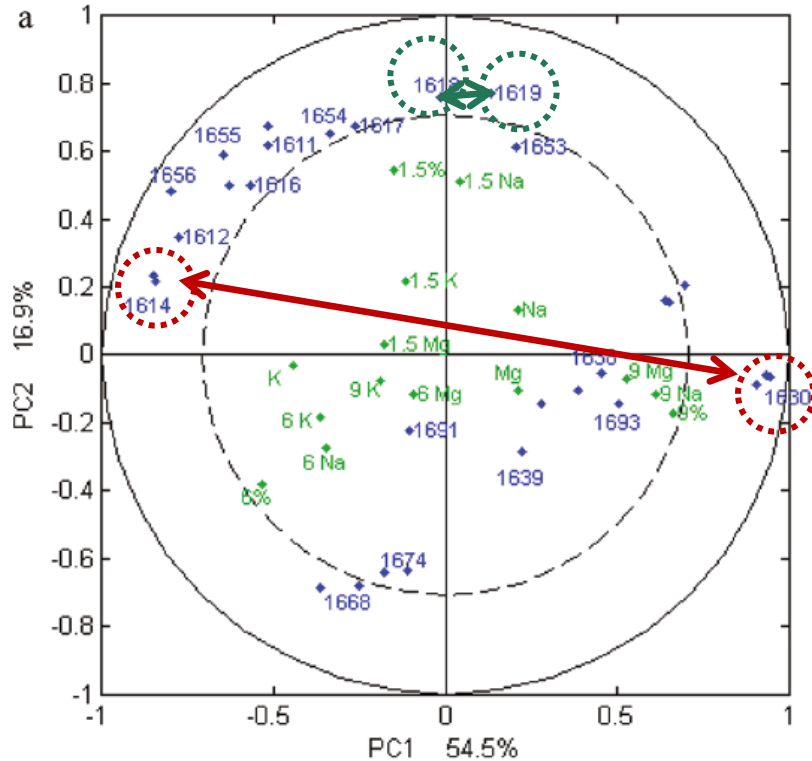
50-100%  
explained variance

importance of a variable

0-50%  
explained variance



## Step 2: PCA of separate regions correlation loading plots: relating to design variables

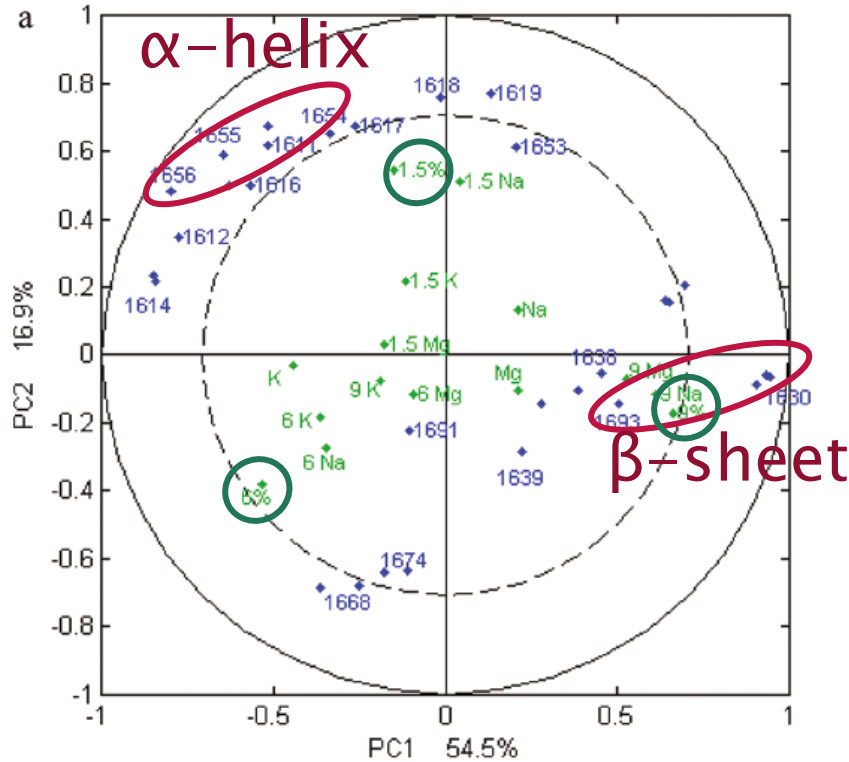


**positive  
correlation**

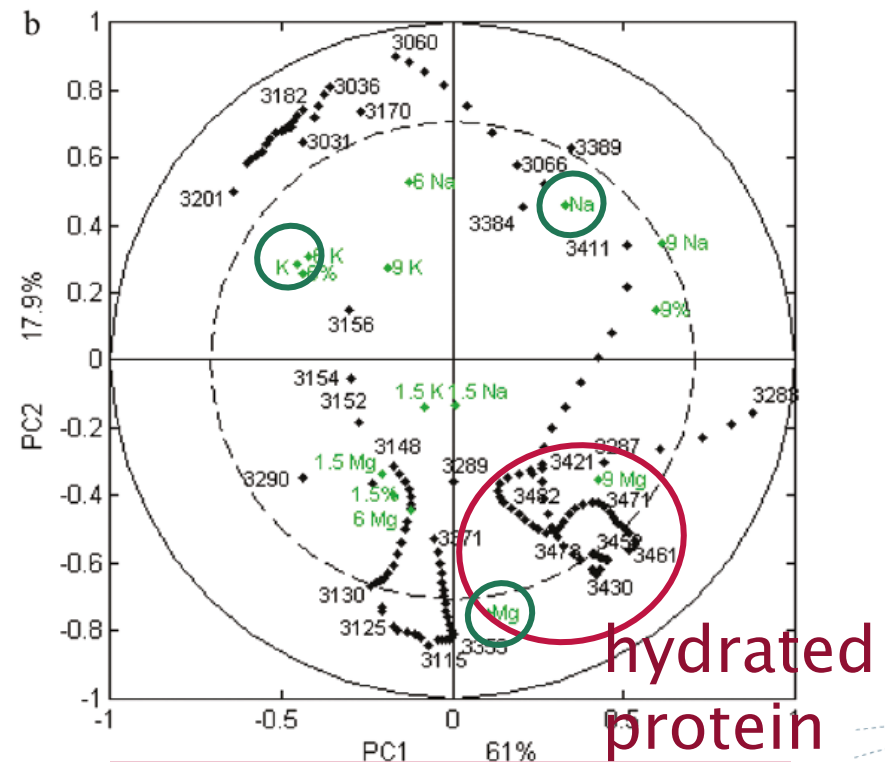
**correlation between  
variables**

**negative  
correlation**

## Step 2: PCA of separate regions correlation loading plots: relating to design variables



protein secondary structure

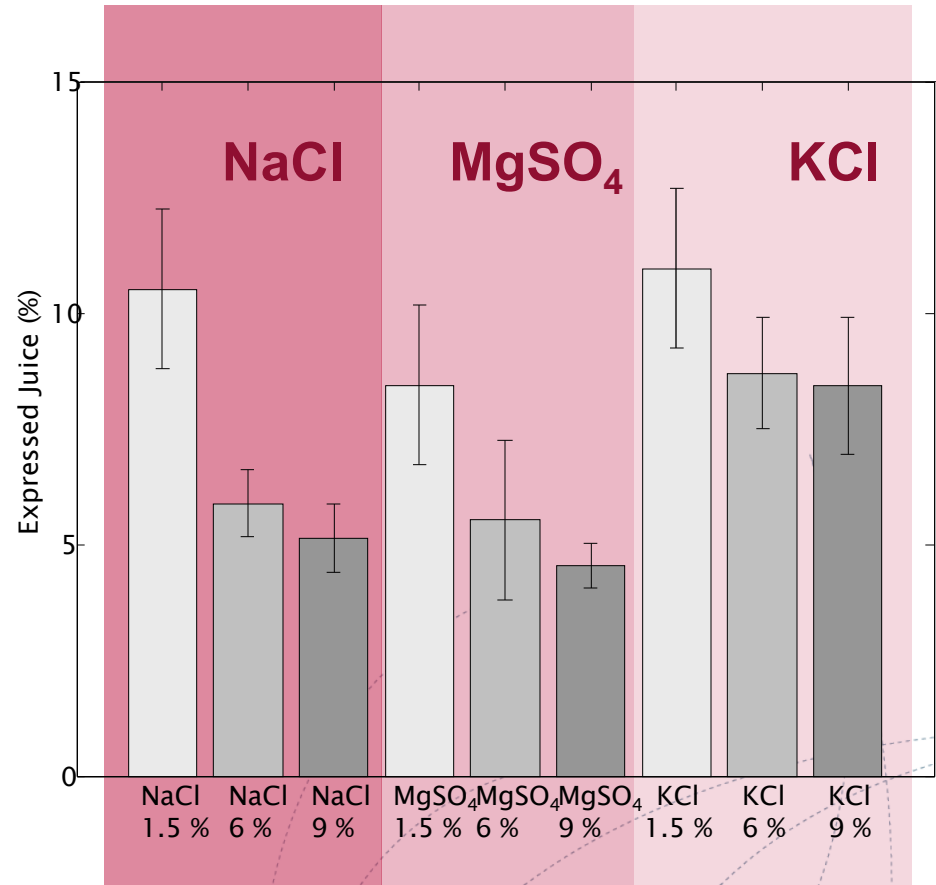


protein water interactions

## Step 4: water holding capacity

WHC  $\uparrow = \downarrow$  juice

- additional measurements (commercial meat samples)
- expressed juice method
- supportig FTIR results:
  - –  $\text{MgSO}_4$  – highest WBC
  - – similar to NaCl
  - – KCl – lowest WBC
  - – different than NaCl



# Characterising protein, salt and water interactions with combined vibrational spectroscopic techniques

## Food Chemistry

Perisic, N., Afseth, N. K., Ofstad, R., Hassani, S. & Kohler, A.  
Characterising protein, salt and water interactions with combined  
vibrational spectroscopic techniques. *Food Chemistry*, 2012, *in press*

# Characterising protein, salt and water interactions with combined vibrational spectroscopic techniques

FTIR microscopy

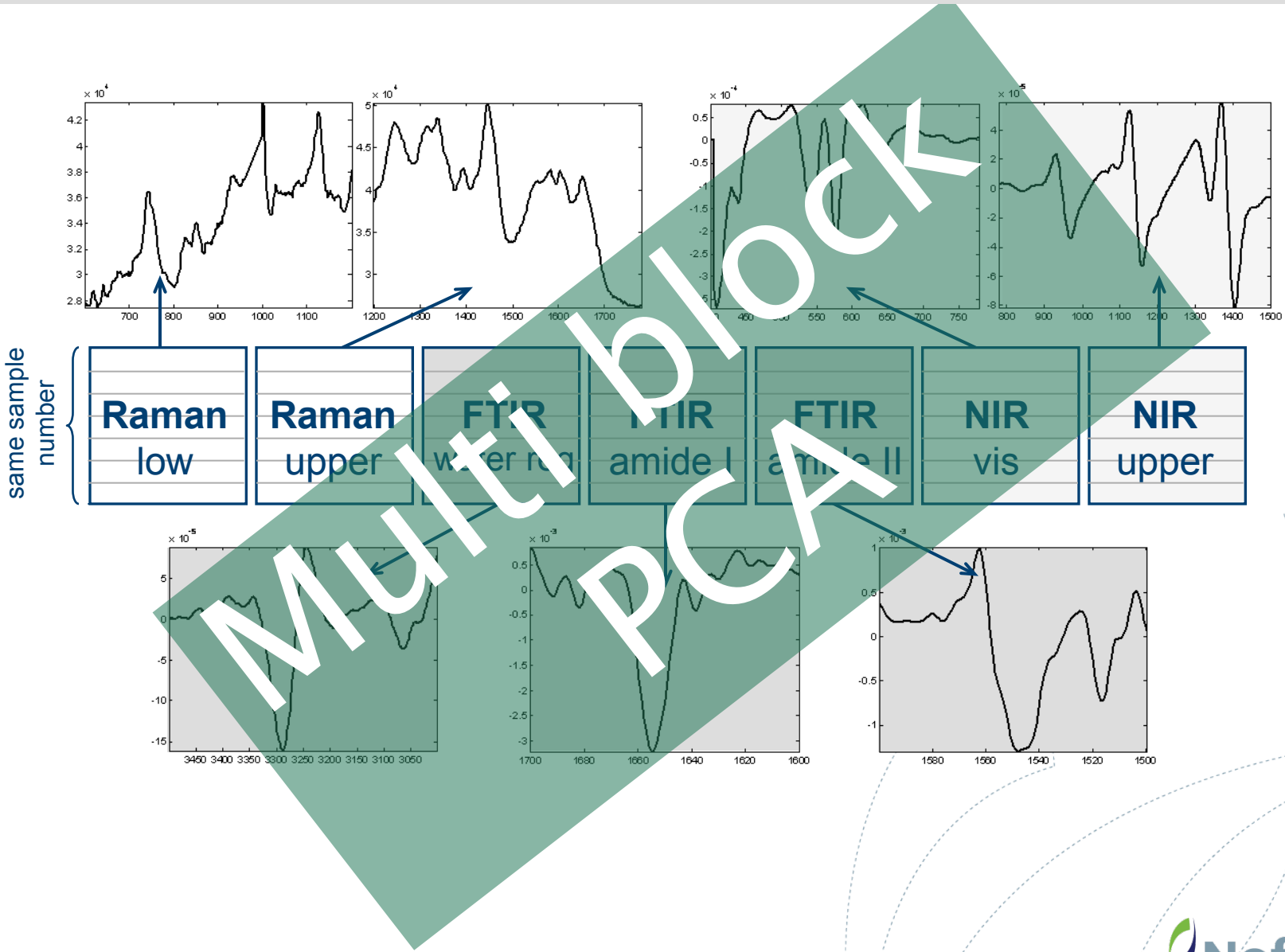
Raman microscopy

NIR spectroscopy

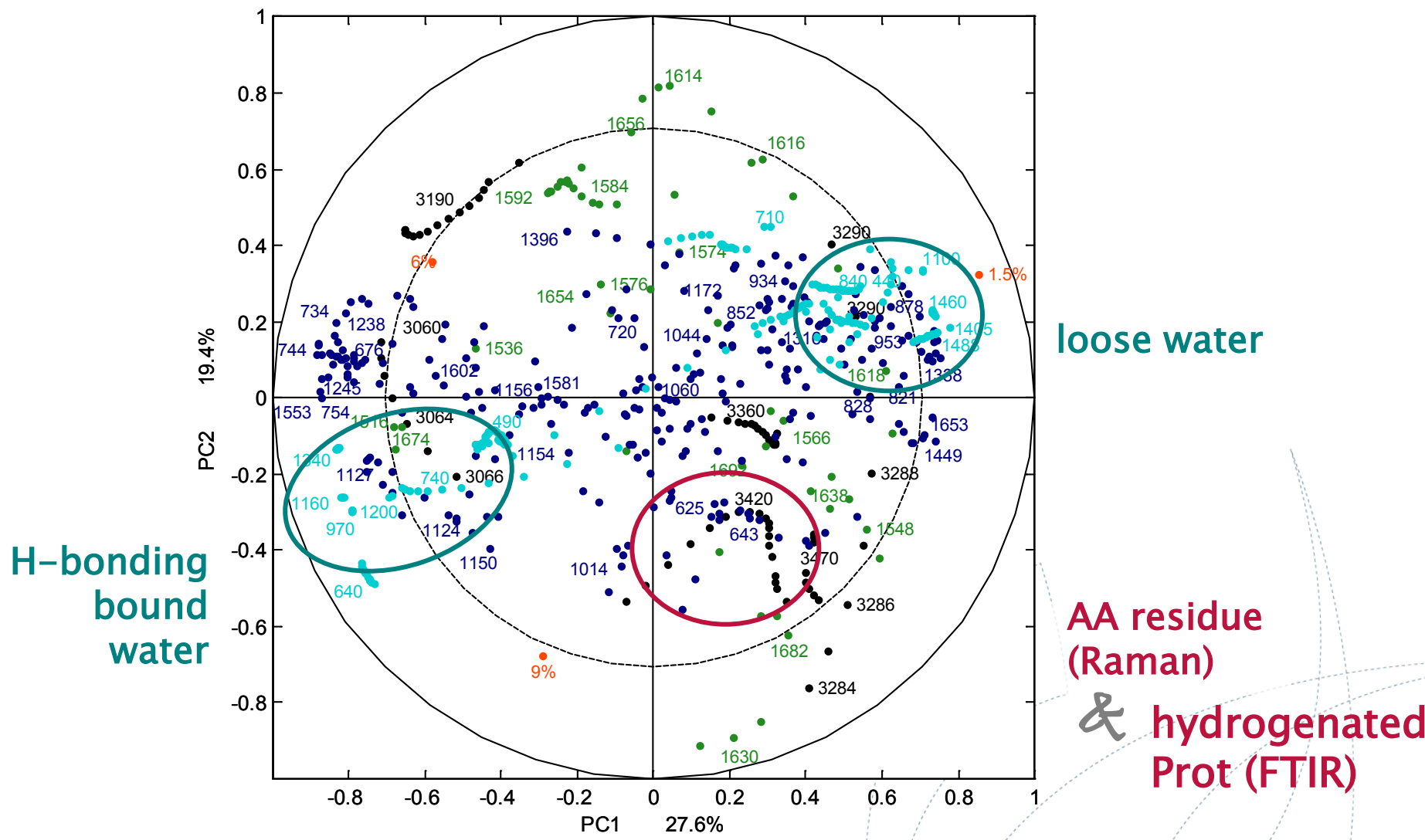
find information  
**characteristic**  
for each method  
&  
**common** between  
methods

# Paper 2

## resulting data set

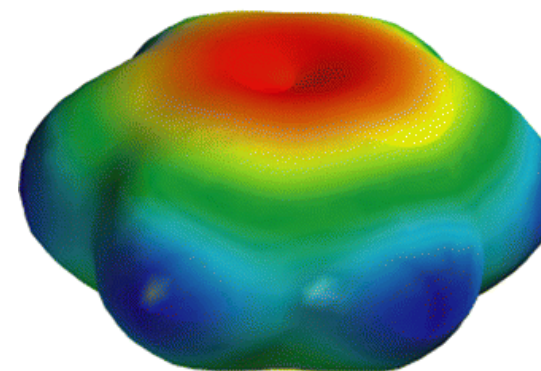
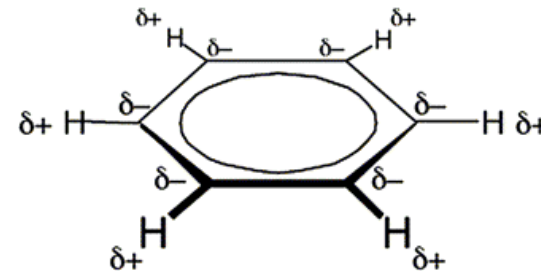
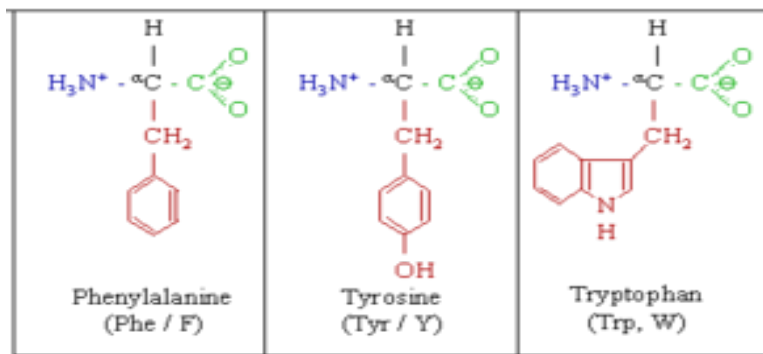


## MB-PCA: correlation loading plot

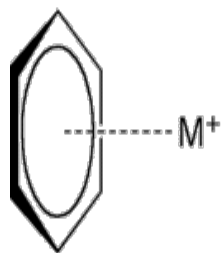


## Cation- $\pi$ interactions: involving aromatic amino acids

### AA residues with aromatic ring



◆ positive charge  
◆ negative charge

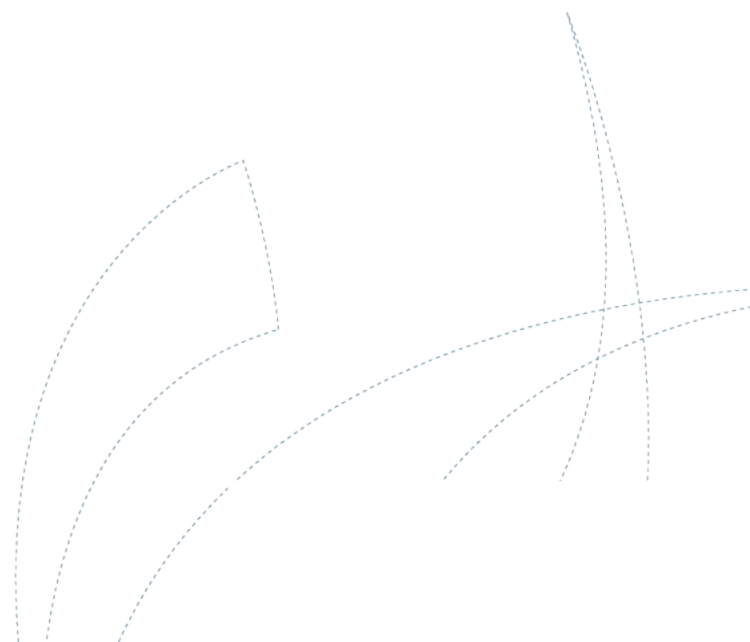
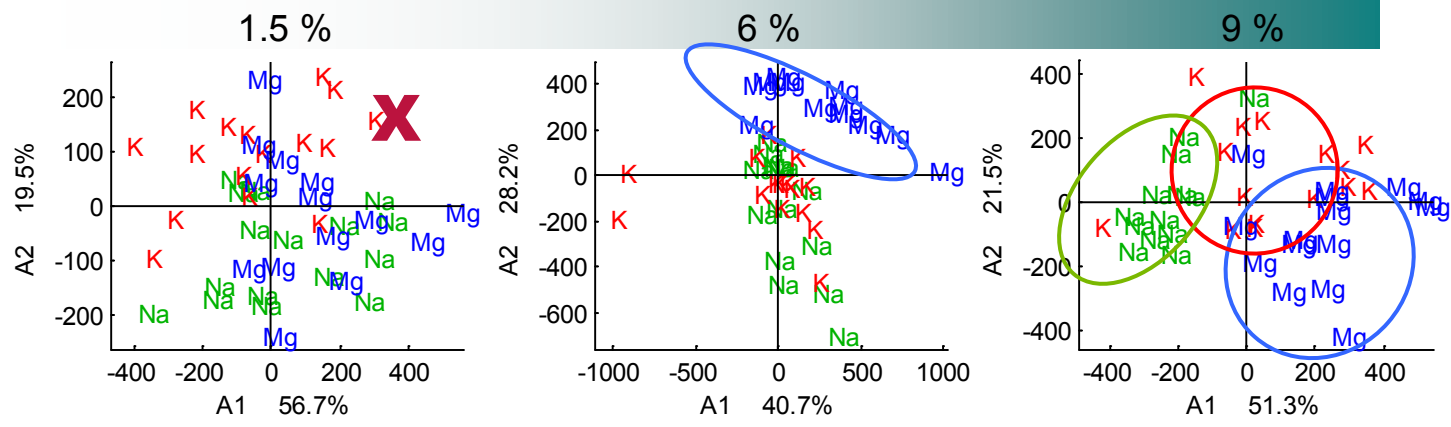


$M^+$	Binding Energy (kcal/mol)	1	2	3
		1 <b>H</b> Hydrogen 1.00794	2 <b>He</b> Helium 4.00260	3 <b>Li</b> Lithium 6.941
$Li^+$	38	4 <b>Be</b> Beryllium 9.012182	5 <b>B</b> Boron 10.811	6 <b>C</b> Carbon 12.011
$Na^+$	28	7 <b>N</b> Nitrogen 14.00643	8 <b>O</b> Oxygen 15.999	9 <b>F</b> Fluorine 18.998403
$K^+$	19	10 <b>Ne</b> Neon 19.992479	11 <b>Na</b> Sodium 22.98976928	12 <b>Mg</b> Magnesium 24.3050
$Rb^+$	16	13 <b>Al</b> Aluminum 26.9815386	14 <b>Si</b> Silicon 28.08558	15 <b>P</b> Phosphorus 30.973762
		16 <b>S</b> Sulfur 32.06	17 <b>Cl</b> Chlorine 35.453	18 <b>Ar</b> Argon 39.948
		19 <b>K</b> Potassium 39.0983	20 <b>Ca</b> Calcium 40.078	21 <b>Sc</b> Scandium 44.955912
		22 <b>Ti</b> Titanium 47.88	23 <b>V</b> Vanadium 50.9415	24 <b>Cr</b> Chromium 51.9961
		25 <b>Mn</b> Manganese 54.938045	26 <b>Fe</b> Iron 55.845	27 <b>Co</b> Cobalt 58.933195
		28 <b>Ni</b> Nickel 58.6934	29 <b>Cu</b> Copper 63.546	30 <b>Zn</b> Zinc 65.38
		31 <b>Ga</b> Gallium 69.723	32 <b>Ge</b> Germanium 72.64	33 <b>As</b> Arsenic 74.9216
		34 <b>Se</b> Selenium 78.96	35 <b>Br</b> Bromine 79.904	36 <b>Kr</b> Krypton 83.80
		37 <b>Rb</b> Rubidium 85.4678	38 <b>Sr</b> Strontium 87.62	39 <b>Y</b> Yttrium 88.90585
		40 <b>Zr</b> Zirconium 91.224	41 <b>Nb</b> Niobium 92.90638	42 <b>Mo</b> Molybdenum 95.94
		43 <b>Ta</b> Tantalum 180.94788	44 <b>Hf</b> Hafnium 178.49	45 <b>Ta</b> Tantalum 180.94788
		46 <b>Hf</b> Hafnium 178.49	47 <b>Ta</b> Tantalum 180.94788	48 <b>W</b> Tungsten 183.84
		49 <b>Re</b> Rhenium 186.207	50 <b>Os</b> Osmium 190.23	51 <b>Ir</b> Iridium 192.222
		52 <b>Pt</b> Platinum 195.084	53 <b>Au</b> Gold 196.96657	54 <b>Hg</b> Mercury 200.59
		55 <b>Cs</b> Cesium 132.90545196	56 <b>Ba</b> Barium 137.327	57 <b>La</b> Lanthanum 138.90547
		58 <b>Ce</b> Cerium 140.12	59 <b>Pr</b> Praseodymium 140.90766	60 <b>Nd</b> Neodymium 144.242
		61 <b>Pm</b> Promethium 144.91288	62 <b>Sm</b> Samarium 150.36	63 <b>Eu</b> Europium 151.964
		64 <b>Gd</b> Gadolinium 157.25	65 <b>Tb</b> Terbium 158.92535	66 <b>Dy</b> Dysprosium 162.5001
		67 <b>Ho</b> Holmium 164.93033	68 <b>Er</b> Erbium 167.259	69 <b>Tm</b> Thulium 168.93032
		70 <b>Yb</b> Ytterbium 173.05468	71 <b>Lu</b> Lutetium 174.967	72 <b>Hf</b> Hafnium 178.49
		73 <b>Ta</b> Tantalum 180.94788	74 <b>W</b> Tungsten 183.84	75 <b>Re</b> Rhenium 186.207
		76 <b>Os</b> Osmium 190.23	77 <b>Ir</b> Iridium 192.222	78 <b>Pt</b> Platinum 195.084
		79 <b>Au</b> Gold 196.96657	80 <b>Hg</b> Mercury 200.59	81 <b>Tl</b> Thallium 204.38
		82 <b>Pb</b> Lead 207.2	83 <b>Bi</b> Bismuth 208.9804	84 <b>Po</b> Polonium 209
		85 <b>At</b> Astatine 210	86 <b>Rn</b> Radon 222	87 <b>Fr</b> Francium 223
		88 <b>Ra</b> Radium 226	89 <b>Ac</b> Actinium 227	90 <b>Th</b> Thorium 232.0377
		91 <b>Pa</b> Protactinium 231.03688	92 <b>U</b> Uranium 238.02891	93 <b>Np</b> Neptunium 237.04817
		94 <b>Pu</b> Plutonium 244.06422	95 <b>Am</b> Americium 243.06138	96 <b>Cm</b> Curium 247.07035
		97 <b>Bk</b> Berkelium 247.07035	98 <b>Cf</b> Californium 251.0832	99 <b>Es</b> Einsteinium 252.0832
		100 <b>Fm</b> Fermium 257.1035	101 <b>Mendelevium</b> 258.1035	102 <b>Nobelium</b> 259.1035
		103 <b>Lr</b> Lawrencium 260.1035	104 <b>Rf</b> Rutherfordium 261.1035	105 <b>Db</b> Dubnium 262.1035
		106 <b>Sg</b> Seaborgium 263.1035	107 <b>Bh</b> Bohrium 264.1035	108 <b>Hs</b> Hassium 265.1035
		109 <b>Mt</b> Meitnerium 266.1035	110 <b>Ds</b> Darmstadtium 267.1035	111 <b>Rg</b> Roentgenium 268.1035
		112 <b>Cn</b> Copernicium 269.1035	113 <b>Nh</b> Nihonium 270.1035	114 <b>Fl</b> Flerovium 271.1035
		115 <b>Mc</b> Moscovium 272.1035	116 <b>Lv</b> Livermorium 273.1035	117 <b>Ts</b> Tennessine 274.1035
		118 <b>Og</b> Oganesson 277.1035	119 <b>Uue</b> Ununennium 278.1035	120 <b>Uub</b> Unbinilium 279.1035

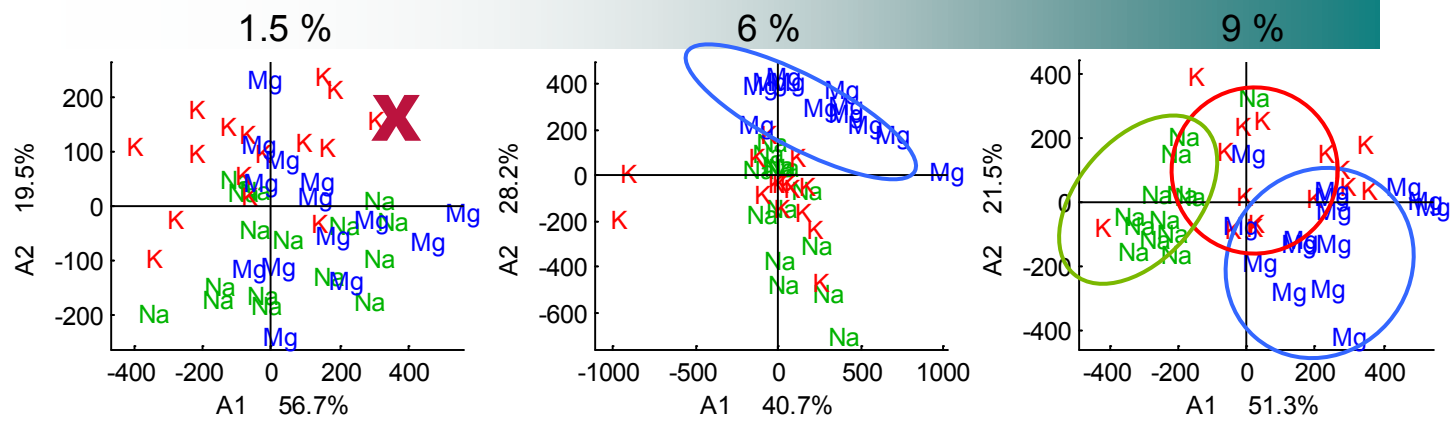
Gas phase binding energies ( $-\Delta H$ ) for alkali metals to benzene.



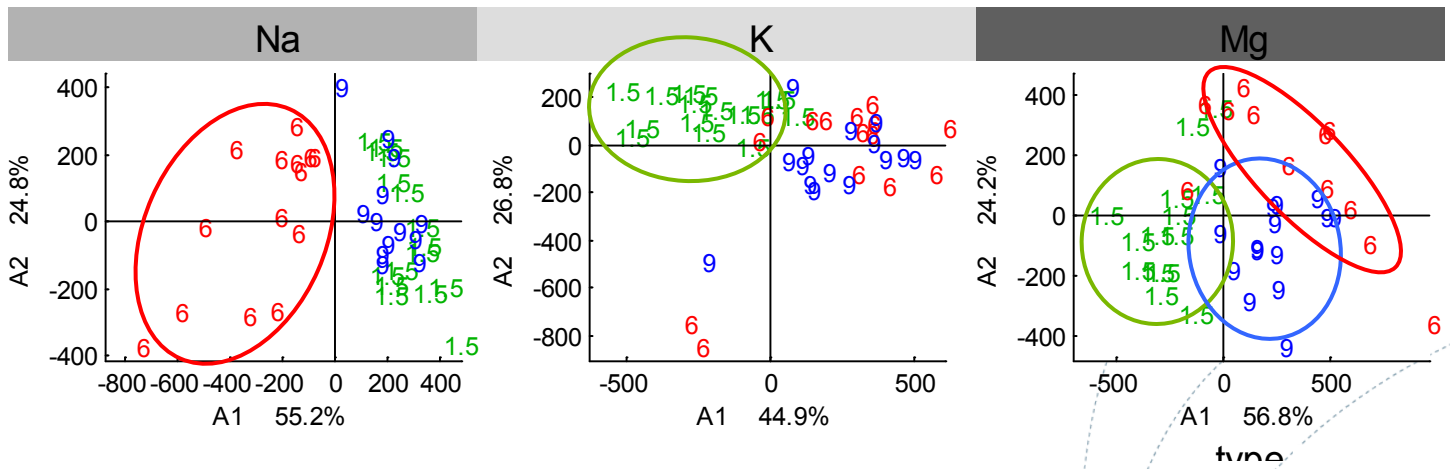
## PCA: Raman AA residue bands



## PCA: Raman AA residue bands



conc.



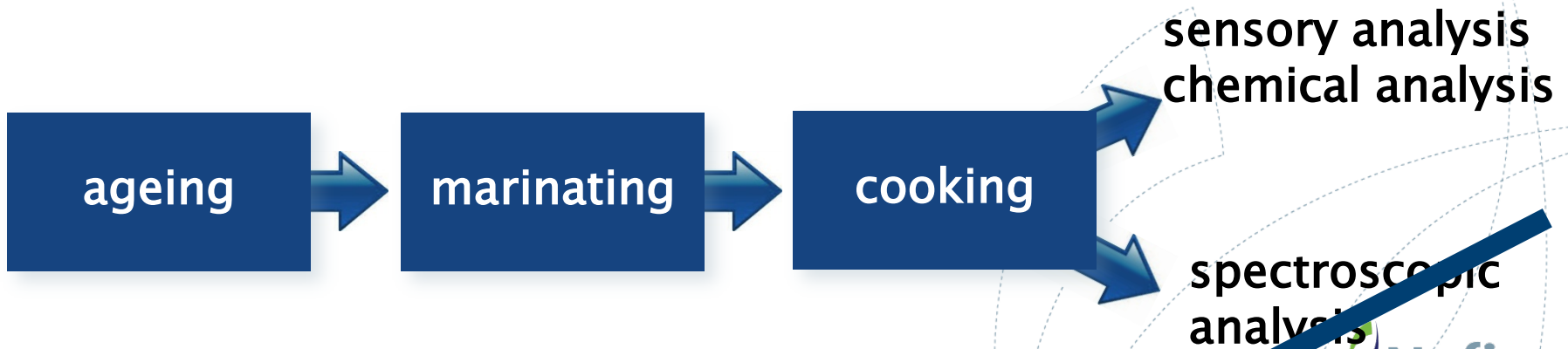
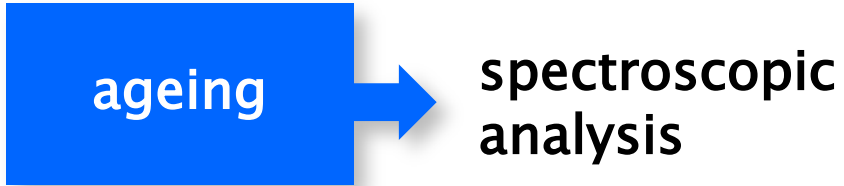
## Characterizing salt substitution in beef meat processing by vibrational spectroscopy and sensory analysis

### Meat Science

Perisic, N.; Afseth, N. K.; Ofstad, R.; Narum, B.; Kohler, A.;  
Characterizing salt substitution in beef meat processing by  
vibrational spectroscopy and sensory analysis

### Experiment 2

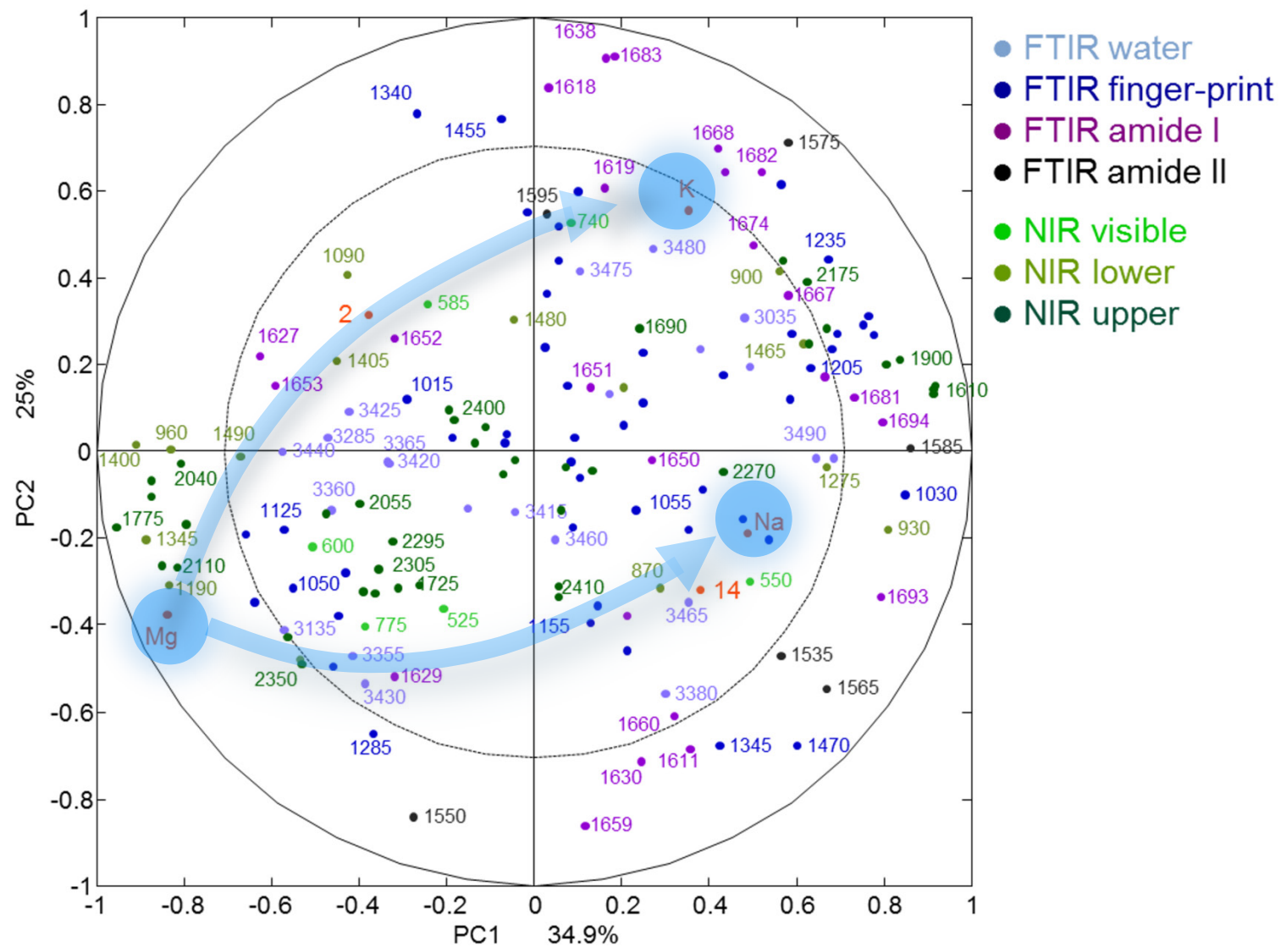
- 3 salts – NaCl / KCl / MgSO<sub>4</sub>
- 1 concentrations – 5,5 % (in marinade)
- 4 animals (beef)
- heat treatment
- ageing
- 2 spectroscopic methods – FTIR, NIR
- sensory analysis





# Paper 3

## spectroscopic analysis



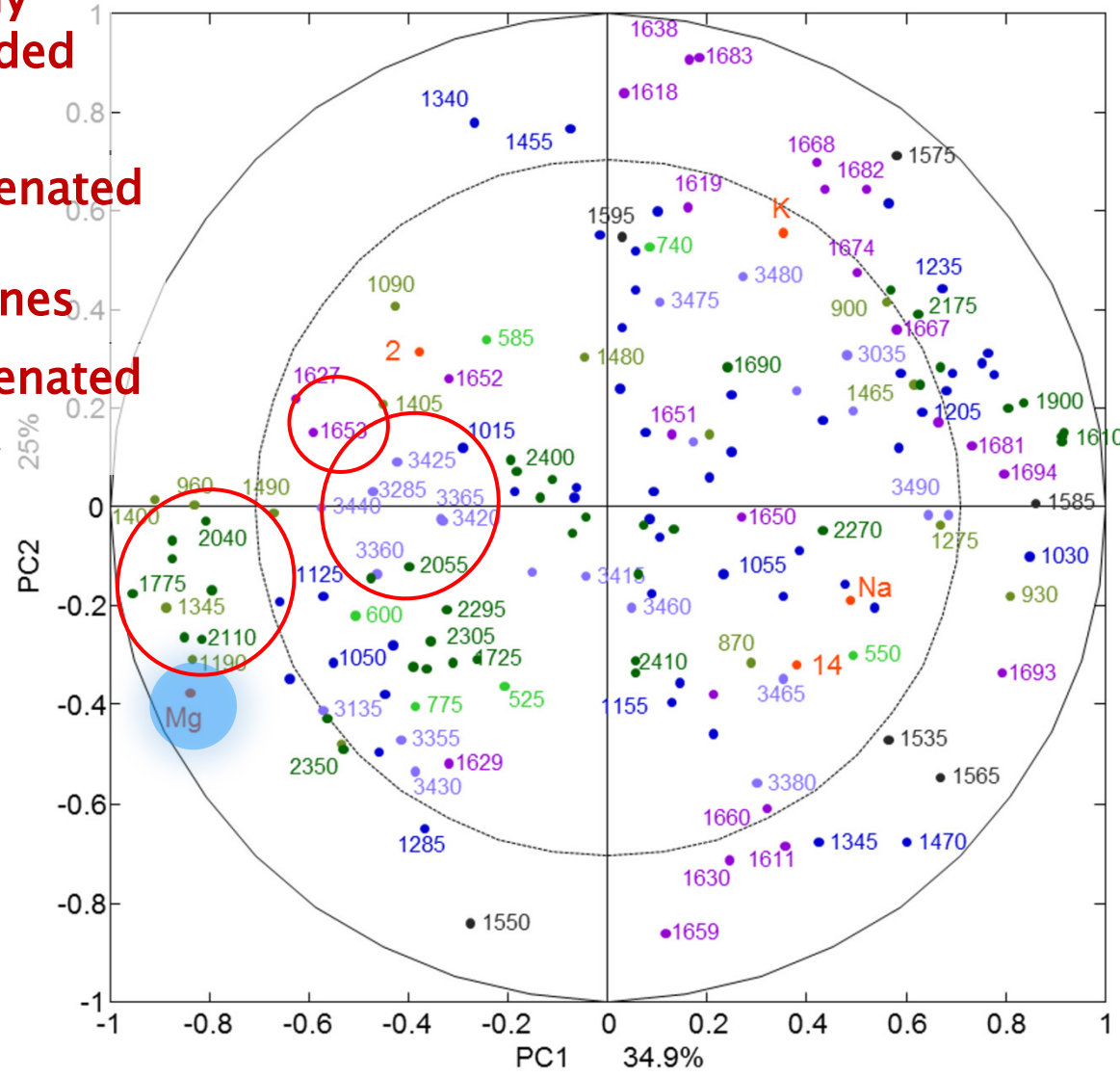
# Paper 3

## spectroscopic analysis

strongly  
H-bonded  
water

hydrogenated  
protein  
backbones

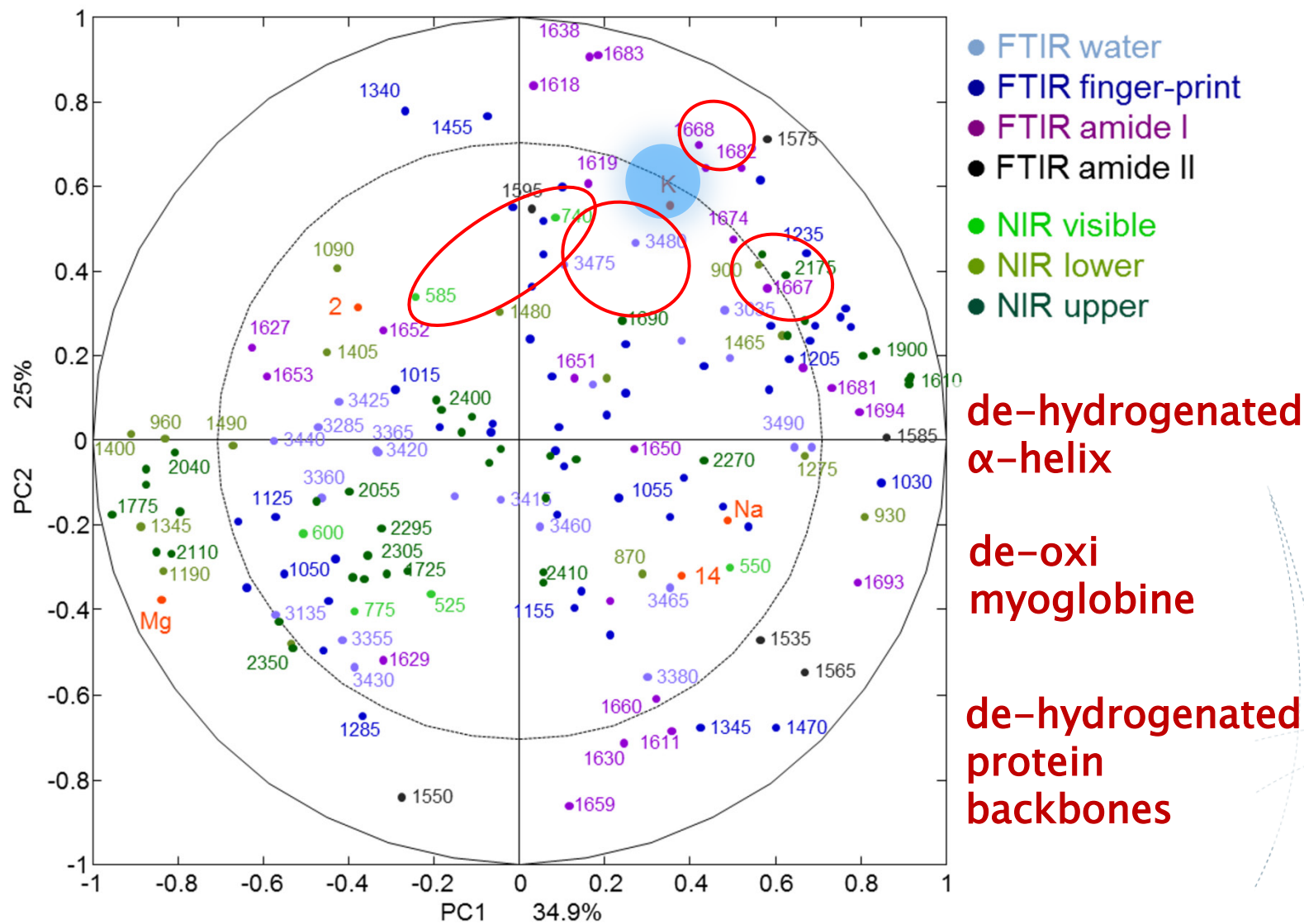
hydrogenated  
 $\alpha$ -helix

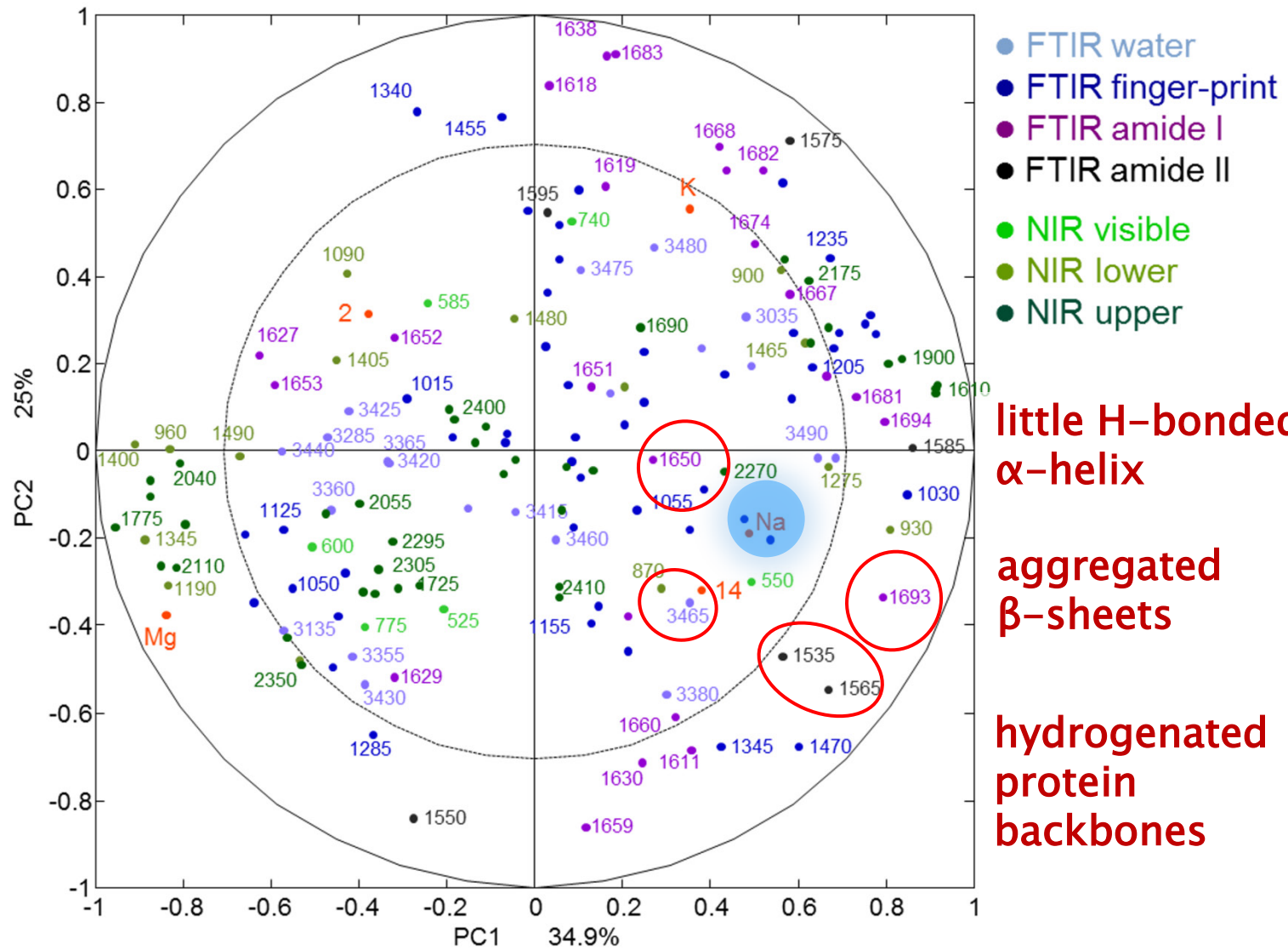


- FTIR water
- FTIR finger-print
- FTIR amide I
- FTIR amide II
- NIR visible
- NIR lower
- NIR upper





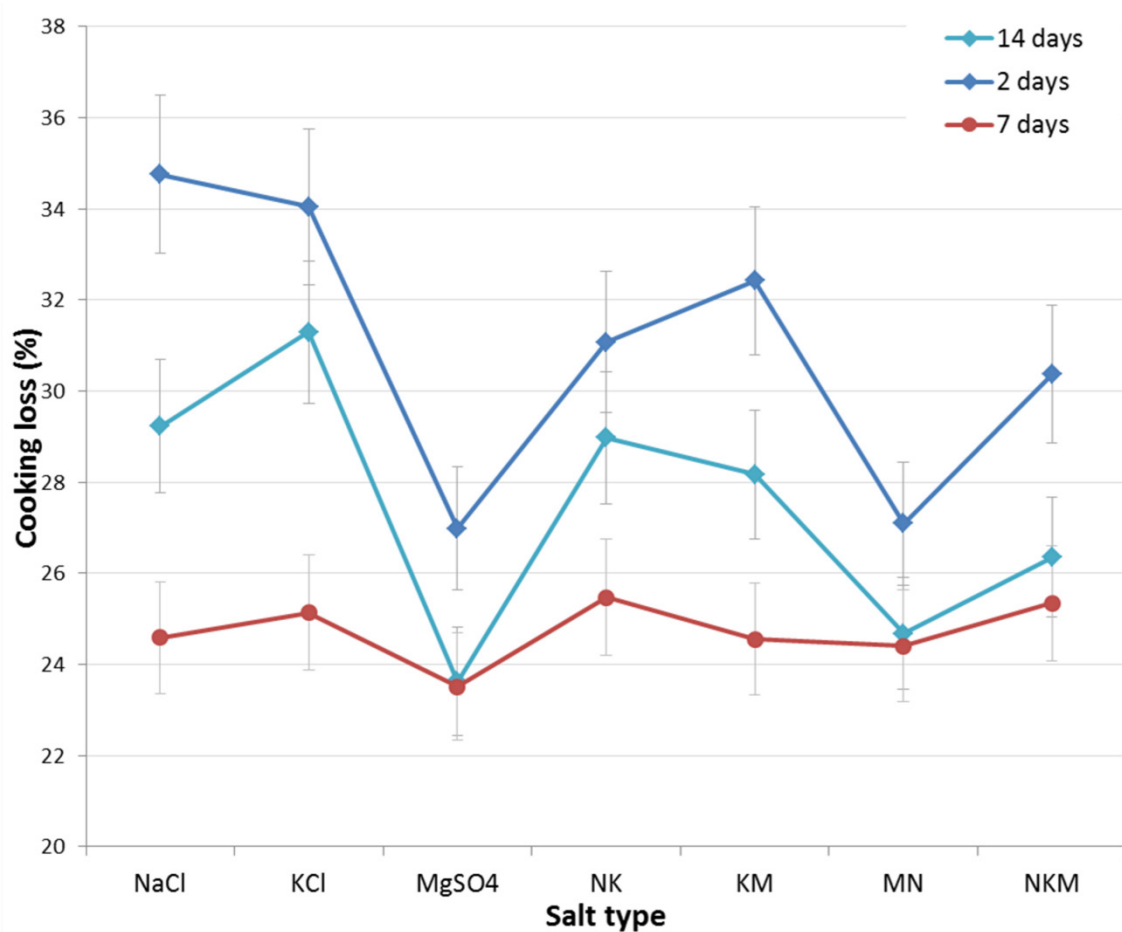




little H-bonded  
 $\alpha$ -helix

aggregated  
 $\beta$ -sheets

hydrogenated  
protein  
backbones



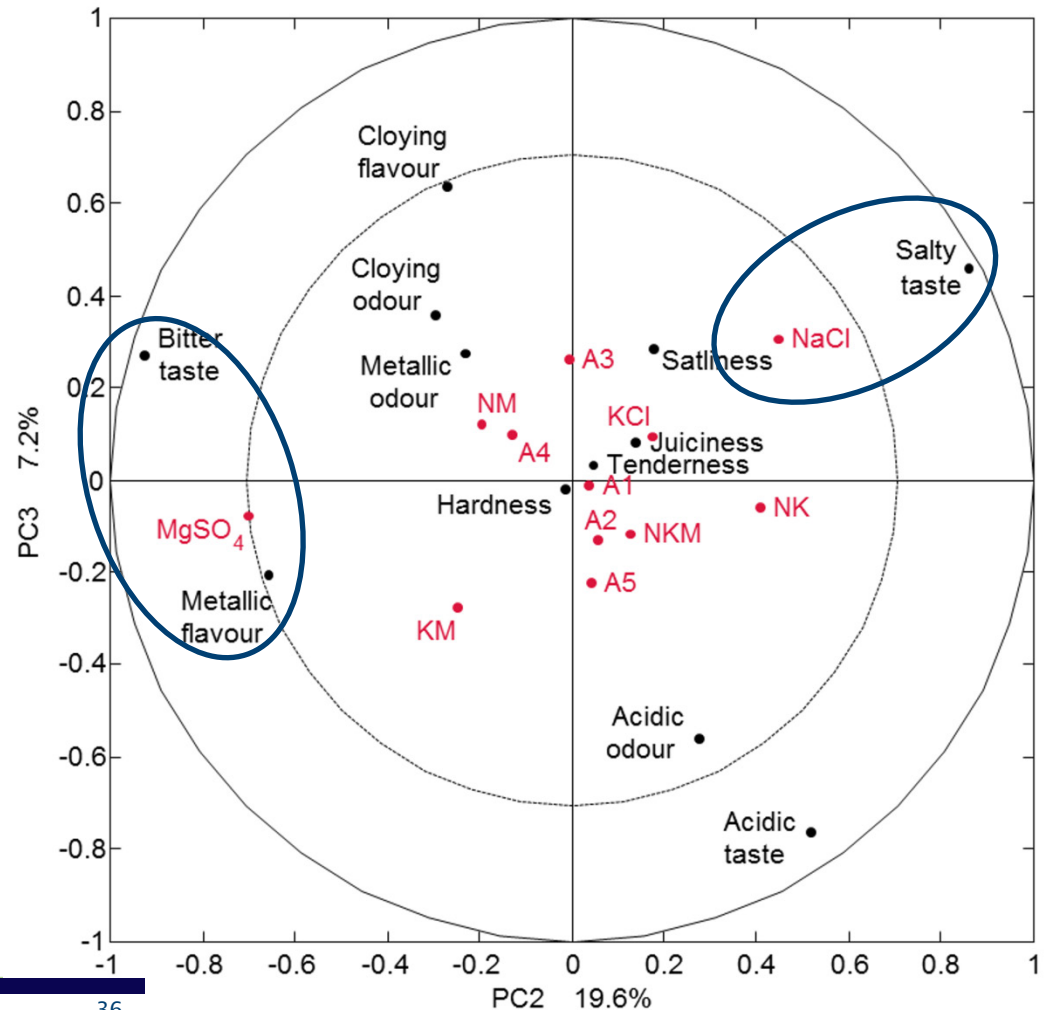
sour, salty, bitter, cloyic, metallic

hardness, tenderness, fetness, juiciness

sour, cloyic, metallic



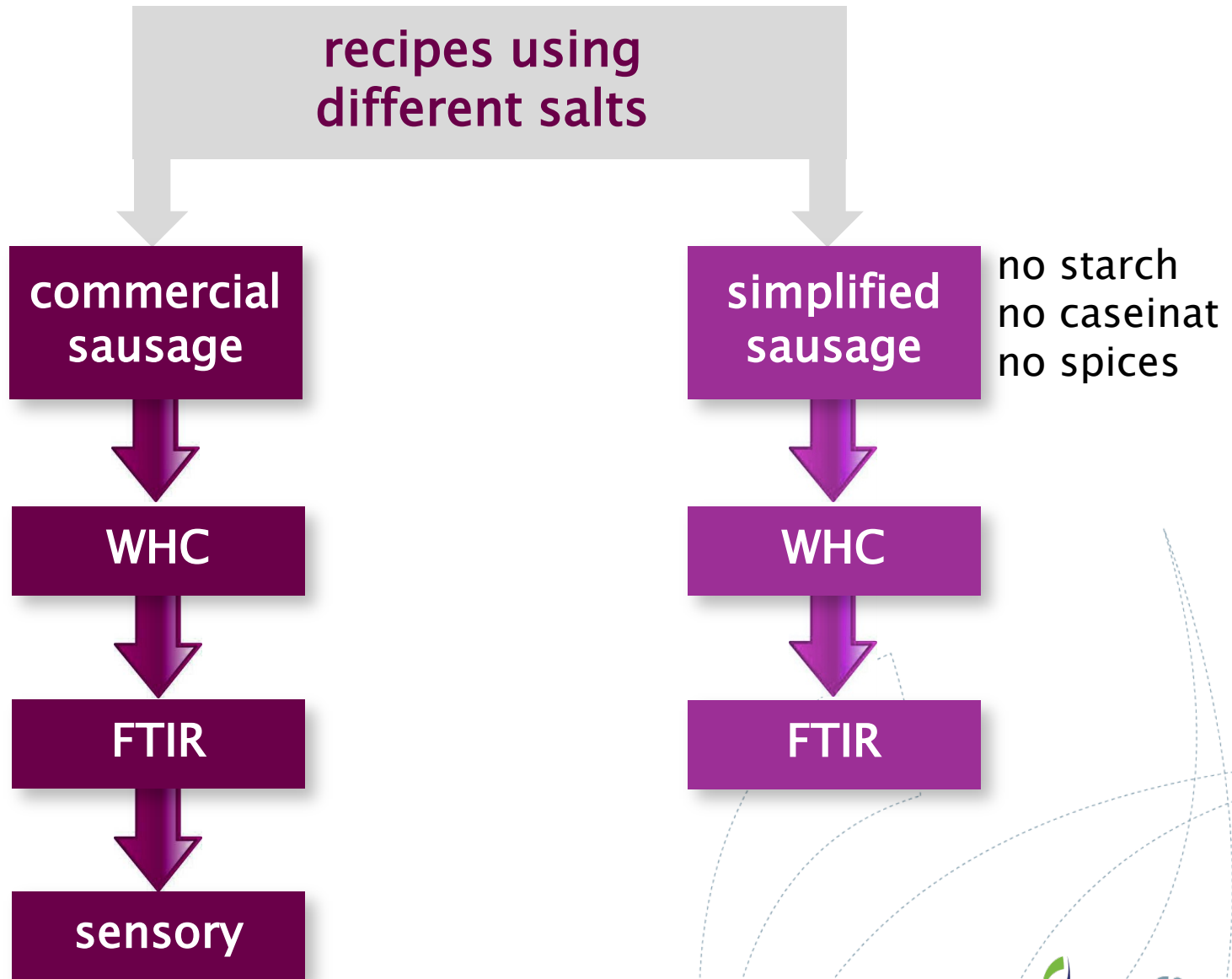
- Evaluated sensory attributes
- Experimental design variables



## Linking structural changes and sensory properties in meat products subjected to salt reduction and salt substitution: a combined FTIR imaging and sensory study

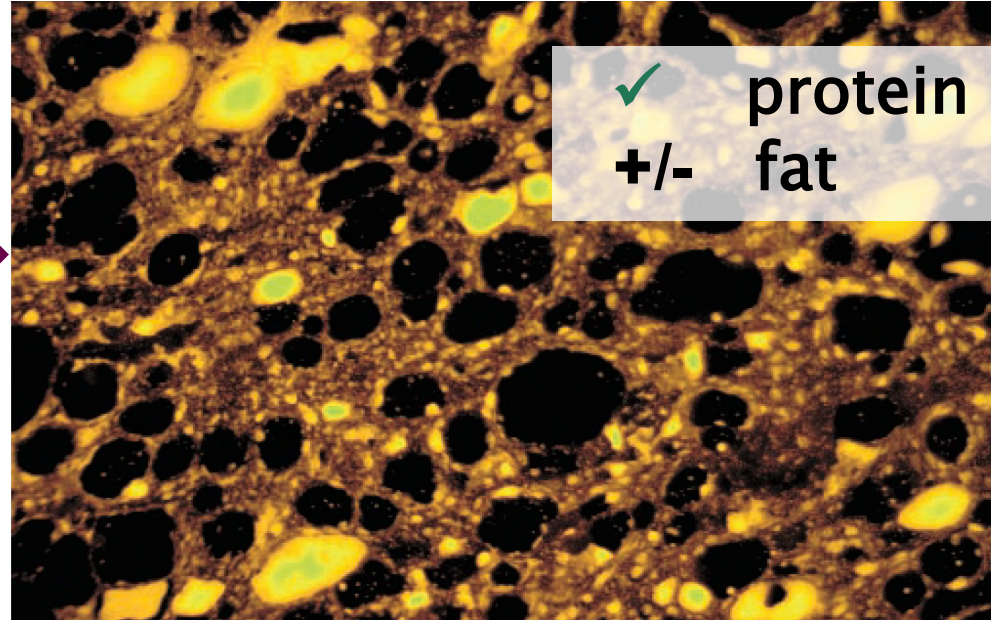
### LWT – Food Science and Technology

Perisic, N.; Afseth, N. K.; Ofstad, R.; Scheel, J.; Kohler, A.; Linking structural changes and sensory properties in meat products subjected to salt reduction and salt substitution: a combined FTIR imaging and sensory study





staining



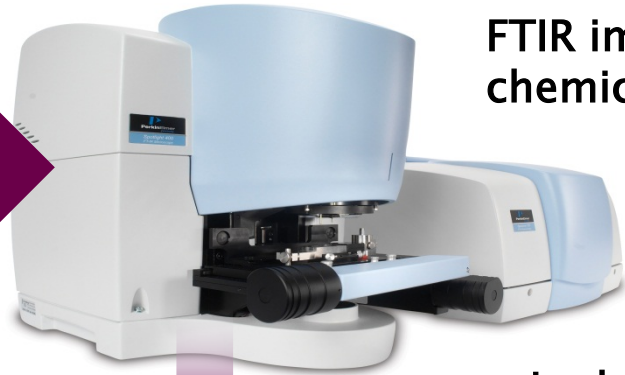
✓ protein  
+/- fat

1 sample = 1-few components

Kohler, A.; Høst, V.; Enersen, G.; Ofstad, R., Identification of fat, protein matrix, and water/starch on microscopy images of sausages by a principal component analysis-based segmentation scheme. *Scanning* 2003, 25, 109–115.

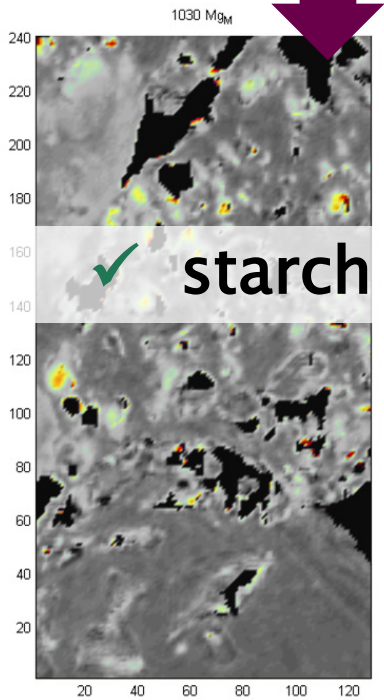
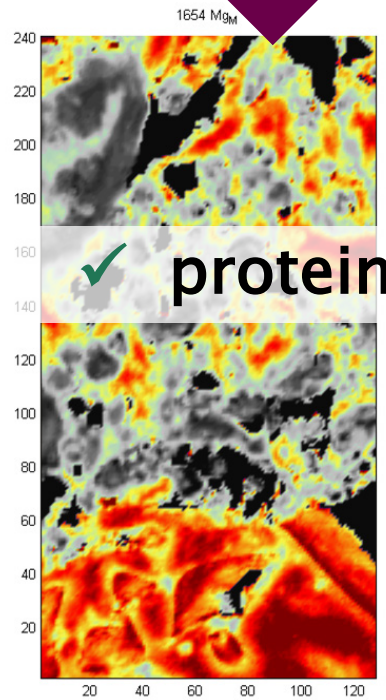
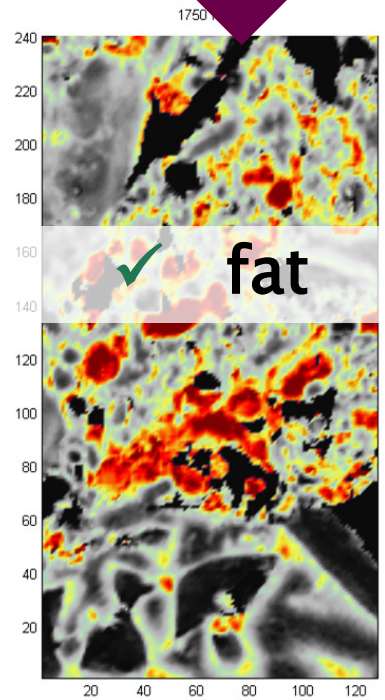
# Paper 4

## background



FTIR imaging  
chemical imaging

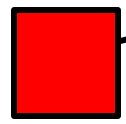
single sample





# Paper 4

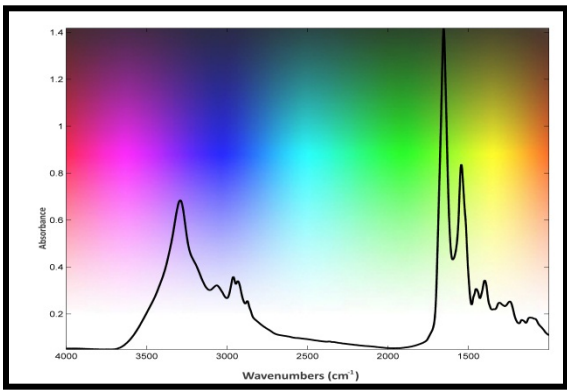
## background



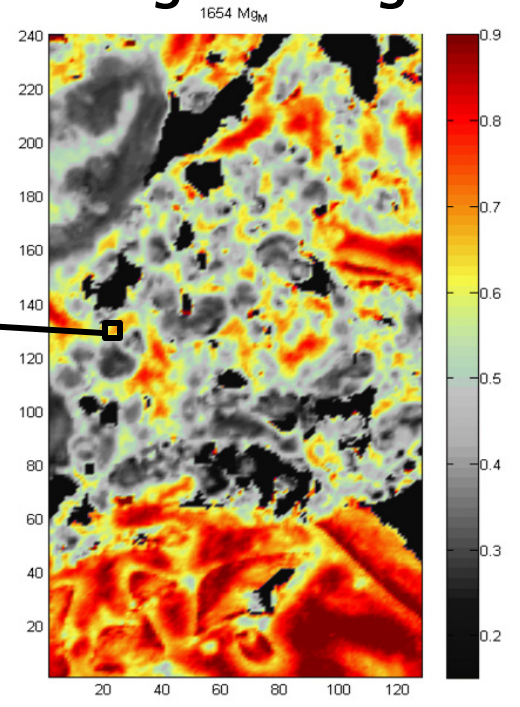
single pixel = single frequency



digital image



single pixel = FTIR spectrum  
= many frequencies  
= many chemical comp.



FTIR image

**thin cross  
sections**





**homogeneity**

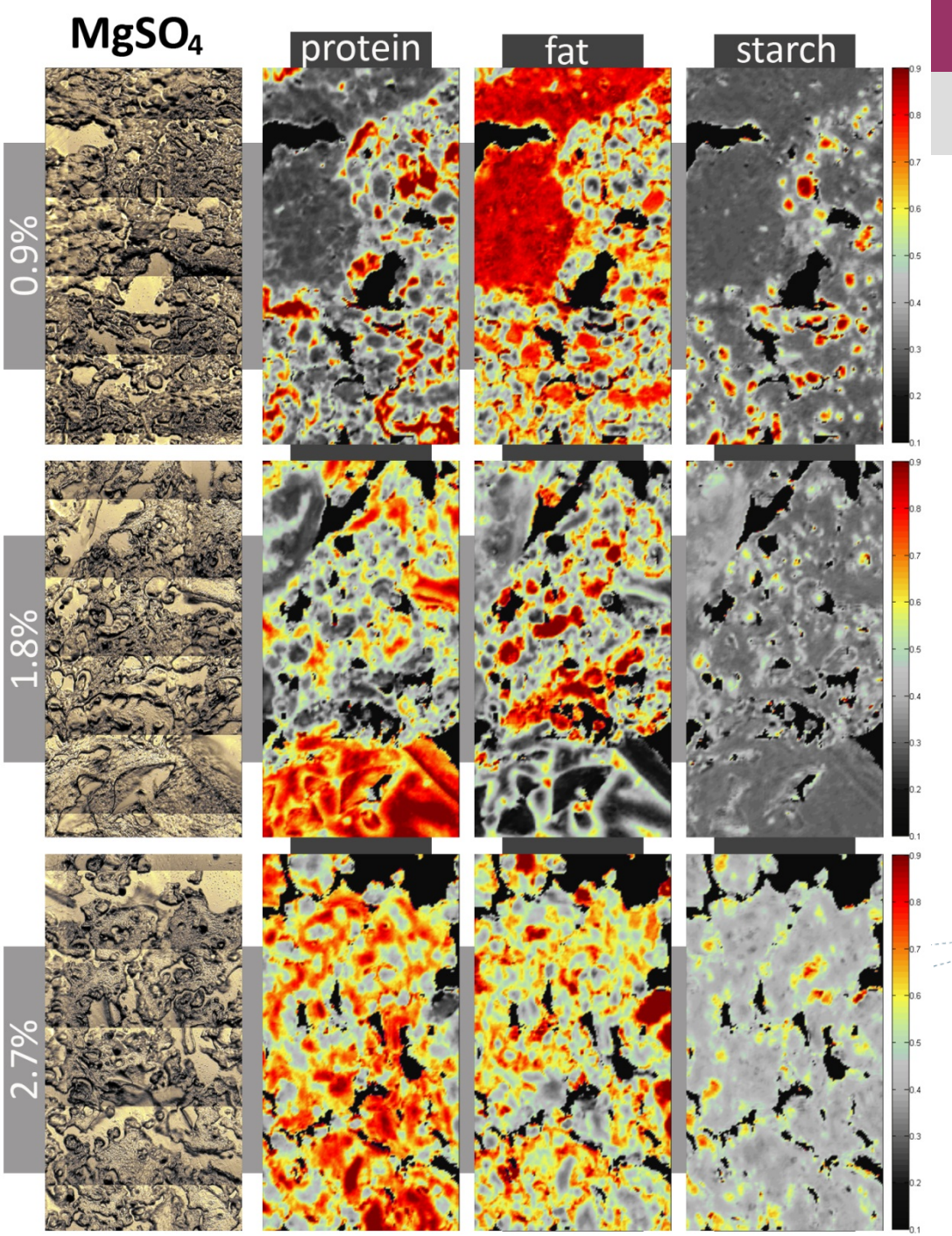


homogeneity



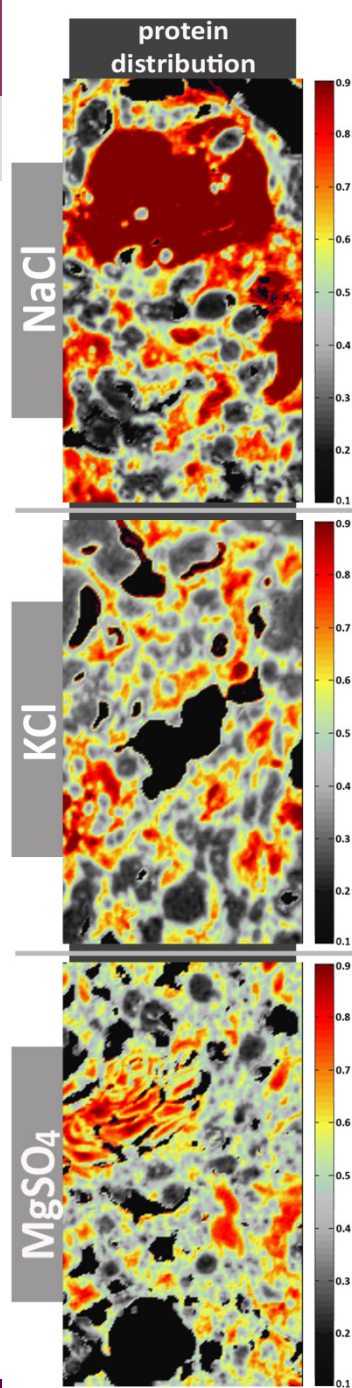
## FTIR imaging results

homogeneity



# Paper 4

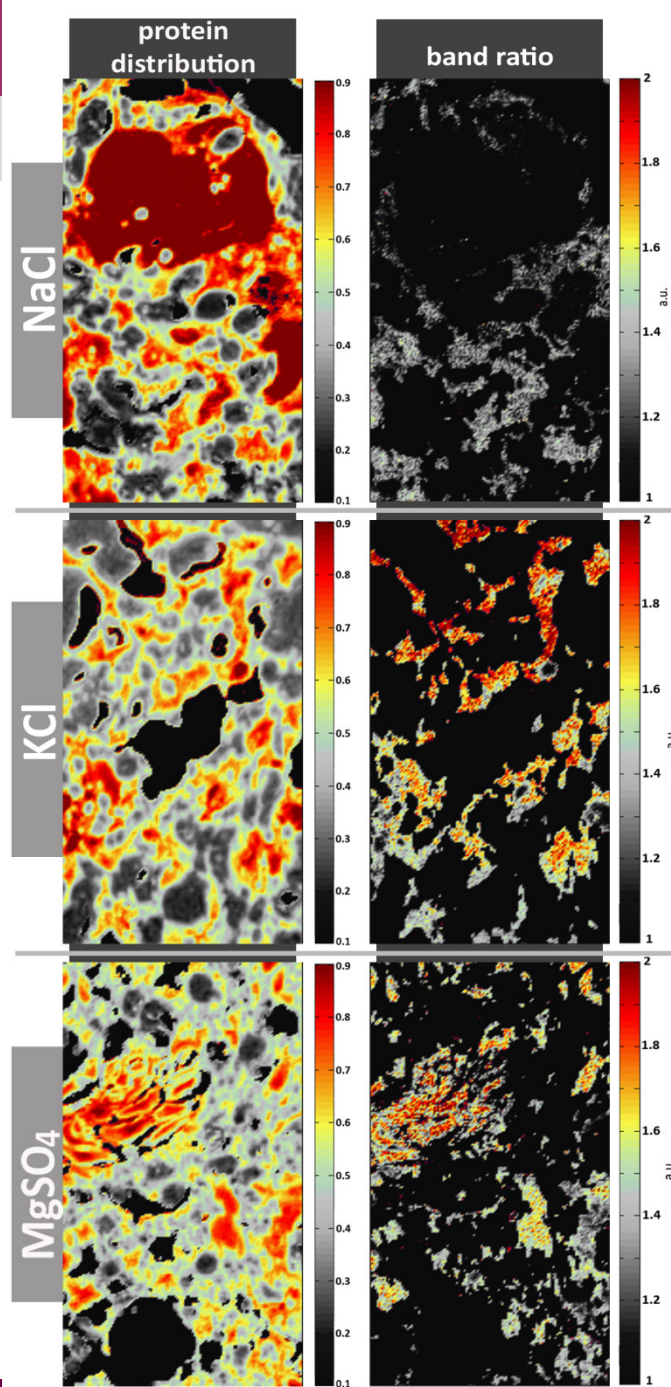
## FTIR imaging results

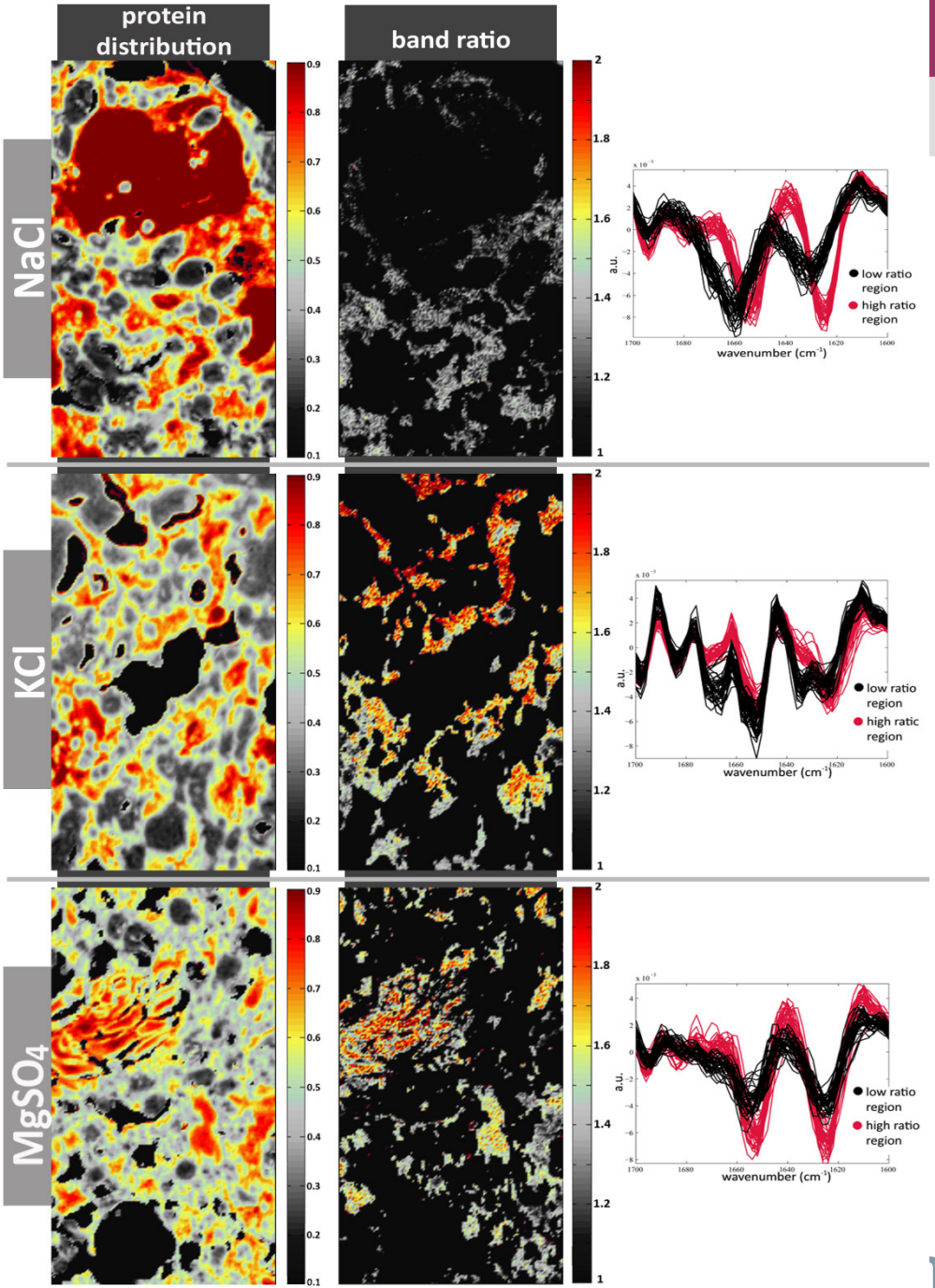


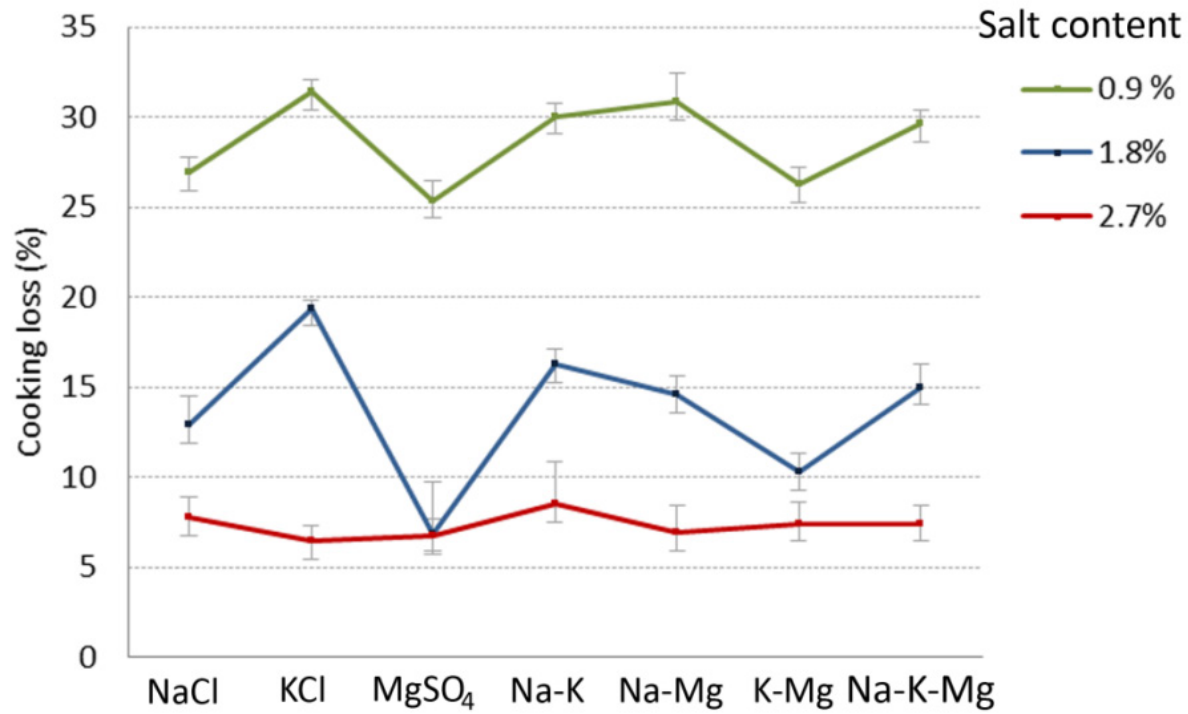


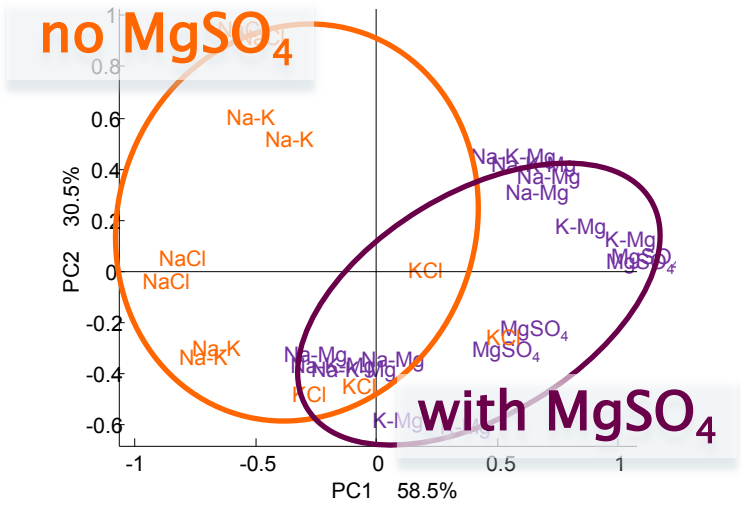
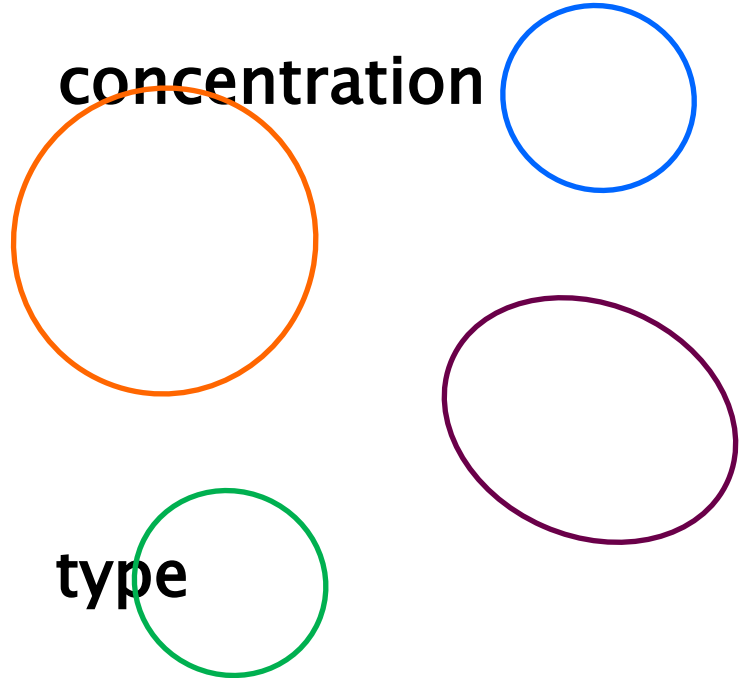
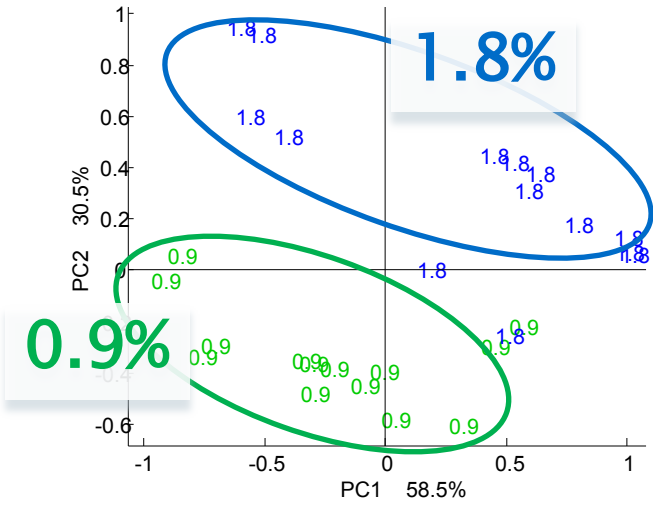
# Paper 4

## FTIR imaging results









salt concentration > salt type

intact meat – salts alone cause only partial denaturation

minced+cooked meat – strong denaturation

– large differences between salts

$\text{MgSO}_4$  – boldly different effect

$\text{MgSO}_4$  – increases WHC  
– hydrogenates proteins

KCl – decreases WHC  
– de-hydrogenates proteins

} from meat model system  
to sausages

only affected by concentration – not salt type

higher concentration – desired texture  
for all salts

$\text{MgSO}_4$  – negative effect on taste  
– when used in low portion – acceptable

# Summary & conclusions

- KCl**
- chemically similar to NaCl
  - bad for increasing WHC
  - bad for taste (high portions)
  - bad for sausage homogeneity

- MgSO<sub>4</sub>**
- chemically different NaCl
  - good/similar effect on WHC as NaCl
  - good for sausage homogeneity
  - **bad** for taste (high portions)

**can be used as a substitute to improve  
WHC / texture / homogeneity**

# Thank you!